

INHIBITION OF THE SRC KINASE FAMILY
PATHWAY AS A METHOD OF TREATING HBV INFECTION
AND HEPATOCELLULAR CARCINOMA

5

1. INTRODUCTION

The present invention relates to therapeutic protocols and pharmaceutical compositions designed to target Src family kinases and components of the Src kinase family signal transduction pathways, including HBx activation of Src kinase family signal transduction pathways for the treatment and prevention of hepatitis B virus (HBV) infection and hepatocellular carcinoma (HCC). The present invention relates to therapeutic protocols and pharmaceutical compositions designed to target cytosolic calcium release or calcium-dependent tyrosine kinase, Pyk2, which is the calcium entry point for activation of Src Kinases for the treatment and prevention of HBV infection and hepatocellular carcinoma. The invention also relates to screening assays to identify potential antiviral agents which target HBx-mediated activation of calcium-dependent tyrosine kinases and Src kinase signaling cascades for the treatment of HBV.

2. BACKGROUND OF THE INVENTION

2.1 Hepatitis B Virus

Infection with HBV is an international public health problem of wide proportions. It has been estimated that at least 10% of the population of tropical Africa and Far-East Asia are chronic carriers of the virus (Tiollais et al., 1985, Nature 317:489-495). HBV is a hepatotropic virus whose course of infection can range from inapparent to acute hepatitis and severe chronic liver disease (Tiollais et al., 1985, Nature 317:489-495). Epidemiological studies have estimated that 250 million people are chronic carriers of HBV

and serve as a reservoir for continued infections. Although the mechanism remains obscure, these HBV carriers have more than a 200 fold greater risk for development of hepatocellular carcinoma (HCC) (Beasley et al., 1981, Lancet 2:1129-1133).

HBV is a DNA-containing para-retrovirus that replicates by reverse transcription but comprises a separate family of viruses from retroviruses, known as hepadnaviruses. Human HBV is the prototype virus in a family that all possess a similar viral architecture and genetic arrangement, although only infection with the mammalian hepadnaviruses HBV (Tiollais et al., 1985, supra), woodchuck hepatitis B virus (WHV) (Popper et al., 1987, Proc. Natl. Acad. Sci. 84:866-870), and possibly ground squirrel hepatitis B virus (GSHV) (Marion et al., 1986, Proc. Natl. Acad. Sci. 83:4543-4546; Seeger et al., 1991, J. Virol. 65:1673-1679) cause both acute and chronic active hepatitis and HCC.

Acute hepatitis following a primary infection with HBV is usually self-limited in adults and often asymptomatic. Following acute hepatitis, 80-90% of infected adult individuals will clear viral antigens from liver and blood, resulting in clinical recovery and immunity to reinfection (Kumar et al., 1992, Basic Pathology, Fifth Edition (Philadelphia: W.B. Saunders Company)). However, 5-10% of individuals do not resolve the primary infection, instead developing a persistent hepatic infection (Ganem and Varmus, 1987, Ann. Rev. Biochem. 56:651-693). Chronic carriers represent a minority outcome following HBV infection, but constitute the majority of cases of HBV-related morbidity and mortality. Infection of infant and newborns results in a high carrier rate (approximately 90%), in contrast to infection of adults. Chronic carriers serve as the reservoir from which HBV is spread both horizontally (through blood and sexual contact) and vertically (from carrier mothers to

newborns). Furthermore, chronic HBV infection frequently results in premature death from hepatic cirrhosis and liver failure (Ganem et al., 1987, Ann. Rev. Biochem. 56: 651-693). As previously noted, chronic carriers have a more than
5 200 fold increased risk for development of primary hepatocellular carcinoma (Beasley et al., 1981, Lancet 2:1129-1133). Because infection by HBV strongly correlates with development of HCC, considerable effort has been expended in identifying potential mechanisms for tumorigenicity by HBV (reviewed in Ganem et al. 1987, supra;
10 Robinson, 1994, Ann. Rev. Med. 45:297-323; Rogler, 1991, Curr. Top. Micro. Immunol. 168:103-140). However, no clear mechanism has been described for the association between HCC and infection with HBV.

There are currently very limited therapeutics available
15 for the treatment of HBV infection. Anti-HBV vaccines are currently being used to prevent HBV infection. However, the efficacy of these vaccines to treat chronic HBV infection and the availability of these vaccines to treat this worldwide health problem remains to be determined. Therefore, the need
20 for an effective anti-HBV therapeutic still exists today.

2.2 HBx

The HBx protein is encoded by one of the four conserved open reading frames of the HBV genome. The L(+) (coding)
25 strand encodes the four conserved open reading frames (ORFs) and codes for all the viral proteins (Ganem et al., 1987, supra). Four mRNAs have been identified. A 2.4 kb preS1 mRNA encodes the large surface antigen (pre-S1) and a 2.1 kb preS2/S mRNA encode the middle (pre-52) and small (major; S) surface antigens (Tiollais et al., 1985, supra). The 3.4 kb
30 pregenome mRNA encodes the precore and core proteins, as well as the polymerase (P). The core protein is the principal structural component of the viral nucleocapsid and possesses

nucleotide binding activity. The P protein, which has RNaseH activity, is the viral reverse transcriptase and the protein primer for synthesis of the L(-) strand (Robinson, 1994, Ann. Rev. Med. 45:297-323). The fourth mRNA is ~0.7 kb in size, and is thought to encode the transcriptional transactivator known as HBx. HBx is a conserved 154 amino acid polypeptide which corresponds to a protein of a molecular weight of ~17 kilodaltons.

The HBx protein is highly conserved within different mammalian HBV serotypes. However, in contrast to the other viral polypeptides, the role of HBx in the HBV life cycle is not yet understood. HBV-infected patient sera indicate that anti-HBx antibodies are produced (Elfassi et al., 1986, Proc. Natl. Acad. Sci. 83:2219-2222; Meyers et al., 1986, J. Virol. 57:101-109), demonstrating that expression of HBx does occur at some stage of HBV infection. HBx protein has also been detected in the livers of patients with chronic hepatitis (Haruna et al., 1991, Hepatol 13:417-421; Katayama et al., 1989, Gastroenterology 97:990-998). Patients testing positive for HBx expression have been found to have increased serum levels of HBV, thereby correlating HBx expression with increased viral replication (Haruna et al., 1991, Hepatol 13:417-421).

The precise role for HBx in the viral infectious process and in the development of HCC remains obscure. There are conflicting reports as to the role of HBx in the viral infectious process and in the development of HCC. It has been reported that there is a correlation between high levels of HBx expression and the development of HCC in transgenic mice. (Kim et al., 1991, Nature 353:317-320; Koike et al., 1994, Hepatol 19:810-819). However, these results remain controversial, as other groups have found no significant liver disease in HBx expressing mice (Balsano et al., 1994,

J. Hepatol. 21:103-109; Dandri et al., 1996, J. Virol. 70; Lee et al., 1990, J. Virol. 64:5939-5947).

Several groups have shown HBx to be a largely if not entirely cytoplasmic protein, although 5-10% of HBx may reside in the nucleus (Doria et al. 1995, EMBO J. 14:4747-4757; Dandri et al. 1996 J. Virol. 70). HBx cannot be found to measurably associate with organelles, membrane vesicles or intermediate filaments, although some preferential accumulation near the cell surface can be observed (Doria et al., 1995, EMBO J. 14:4747-4757). HBx is a weak to moderately strong transcriptional transactivator. HBx has been shown to transactivate transcription of the interferon- β gene (Twu et al., 1987, J. Virol. 61:3448-3453) and of the HBV enhancer (Spandau et al., 1988, J. Virol. 62:427-434). Since those first reports, HBx has been shown to transactivate a wide variety of cellular and viral transcriptional elements (reviewed in Yen, 1996, J. Biomed. Sci. 3:20-30). Activation has been localized to specific binding sites for the transcription factors AP-1 (Benn & Schneider, 1994, Proc. Natl. Acad. Sci. 91; Natoli et al., 1994, Mol. Cell. Biol. 14:989-998; Seto et al., 1990, Nature 344:72-74), AP-2 (Seto et al., 1990, supra), NF- κ B (Lucito & Schneider, 1992, J. Virol. 66:983-991; Mahe et al., 1991, J. Biol. Chem. 266:13759-13763; Su and Schneider, 1996 J. Virol. 70:4558-4566; Twu et al., 1989, J Virol. 61:3448-3453), ATF/CREB (Maguire et al., 1991, Science 252:842-844) and possibly c/EBP (Faktor and Shaul, 1990; Mahe et al., 1991, supra).

HBx does not contain any structural motifs that convincingly suggest a function, such as DNA binding (Lucito & Schneider, 1993 in Animal Viruses, L. Carrasco ed. (NY: Plenum Press) p. 67-80), nor has it been observed to directly bind DNA (Siddiqui et al., 1987, Virol. 169:479-484; Wu et al., 1990 Proc. Natl. Acad. Sci. USA 84:2678-2682). A number

of activities have been ascribed to HBx including an *in vitro* association with p53 (Butel et al., 1996, *Trend Micro.* 4:119-124), an association with the human homolog of a UV-damage DNA repair protein (Lee et al., 1995, *J. Virol.* 69:1107-1174), and an association with a serine protease inhibitory protein (Takada et al., 1994, *Oncogene* 9:341-348). In summary, it appears that the activities of HBx are not limited solely to transcriptional transactivation, and surely other HBx-associated activities will be discovered.

One early model suggested that HBx indirectly stimulates transcription through activation of a protein kinase C (PKC) signaling pathway (Kekule et al., 1993, *Nature* 361:742-745). Many groups report PKC-independent transactivation by HBx (Benn et al., 1996, *J. Virol.* 70:4978-4985; Chirillo et al., 1995, *J. Virol.* 70; Cross et al., 1993, *Proc. Natl. Acad. Sci.* 90:8078-8082; Lucito & Schneider, 1992, supra; Murakami et al., 1994, *Virol.* 199:243-246; Natoli et al., 1994, supra). It was demonstrated that HBx activation of AP-1 and NF- κ B factors occurs by HBx activation of a Ras signal transduction cascade (Benn & Schneider, 1994 supra; Cross et al. supra; Natoli et al., 1994, supra; Su & Schneider, 1996, supra). HBx was shown to stimulate RasGTP complex formation and to establish a cascade linking Ras, Raf, and MAP Kinase, which is essential for HBx activation of AP-1 (Benn & Schneider, 1994, supra) and NF- κ B (Su and Schneider, 1996, supra). However, the mechanism by which HBx stimulates RasGTP complex formation remains to be elucidated. Additional results have also shown that HBx stimulates cellular proliferation in quiescent cells and induces deregulation of cell cycle checkpoint controls in a Ras dependent manner (Benn & Schneider, 1995, *Proc. Natl. Acad. Sci. USA* 92:11215-11219), indicating that activation of Ras by HBx appears to play a central role in defining HBx activities.

3. SUMMARY OF THE INVENTION

The present invention relates to the treatment and prevention of HBV infection by targeting activation of the Src family of kinases. The present invention further relates to the treatment and prevention of HBV infection by targeting activation of cytosolic calcium release and Pyk2-Src signal transduction. The present invention also relates to compounds which inhibit HBx-mediated activation of the Pyk2 tyrosine kinase and Src family of kinases as well as the downstream components of the Pyk2-Src kinase signaling cascade for the treatment of HBV infection.

The Applicants have shown that HBx activation of Src kinases stimulates viral DNA replication, and HBx activates Src kinases by stimulating two related upstream tyrosine kinases known as Pyk2 and p125FAK (FAK). The Applicants have shown that HBx activation of Pyk2, FAK, Src and MAPK signalling, all occur in a calcium-dependent manner in that treatment of cells with calcium chelator (EGTA) or calcium channel poison (BAPTA-AM) specifically blocks HBx stimulation of Pyk2, which is essential for HBx activity. In addition, treatment of cells with cyclosporin A (CsA), a specific inhibitor of mitochondrial voltage-dependent anion channels, which deregulates calcium channels, also impairs HBx stimulation of HBV genomic DNA replication. Thus, the Applicants have demonstrated that HBx functions through a calcium-dependent pathway to stimulate viral DNA replication in cells and Pyk2 signal transduction, which plays a fundamental role in mammalian hepadnavirus replication.

The Applicants have demonstrated that HBx mediated activation of Pyk2-Src kinase signaling cascade is an effective target for HBV anti-viral agents since activation of this pathway is essential for HBV replication. The Applicants have further demonstrated that HBx acts through calcium channels or their regulatory components to sustain

102750-900000050
Sub. 27

HBV replication. Therefore, targeting HBx for the treatment of HBV should result in a highly specific and efficacious method of blocking HBV replication. The Pyk2-Src family of kinases, although host cell gene products, are only activated
5 in proliferating or differentiating cells, and in cells infected by many DNA and tumor viruses. Therefore, targeting the Pyk2-Src kinase transduction pathway for the treatment of HBV infection should result in a therapeutic with a high degree of efficacy and sufficient specificity with side
10 effects no more toxic than chemotherapeutics currently used to treat cancer.

The present invention encompasses a variety of techniques and compounds to target the activities of HBx essential for HBV replication. In particular, these include, but are not limited to HBx-mediated activation of the Src
15 kinase family signal transduction pathways for the treatment and prevention of HBV infection. The present invention encompasses the use of known inhibitors of Pyk2 tyrosine kinase signal transduction, in addition to inhibitors of calcium channels and their regulatory components, to treat
20 HBV infection. The invention encompasses the use of known Src inhibitors to treat HBV infection. Examples of such specific inhibitors include, but not limited to: Pyk2 specific tyrosine kinase inhibitors, Src specific tyrosine kinase inhibitors, such as CsK, tyrphostin-derived
25 inhibitors, derivatives of benzylidenemalonitrile, pyrazolopyrimidine PP1, and microbial agents, such as angelmicin B; and competitive inhibitors, such as small phosphotyrosine containing ligands. The invention also encompasses the use of known HBx inhibitors for the treatment of HBV, including, but not limited to, antisense RNAs
30 directed to HBx. The present invention also relates to the use of inhibitors of downstream effectors of Src kinases, including but not limited to, cytoplasmic factors, such as

Ras, Raf, focal adhesion kinase (FAK) and MAPK, and nuclear factors, such as Myc and cyclin-dependent kinases.

In another embodiment of the present invention gene therapy approaches, including dominant-negative mutants, antisense molecules and SELEX RNAs targeted to block Src kinase or HBx gene expression, may be used as a method to treat and prevent HBV infection and HCC. In yet another embodiment of the invention, upstream and downstream components and effectors of the Src kinase family signaling cascade may be targeted by gene therapy approaches to inhibit HBV infection.

The present invention further relates to screening assays to identify compounds which inhibit HBx-mediated activation of the Src kinase signaling pathway and may be used to treat HBV infection and diseases and disorders associated with HBV infection. The present invention also relates to screening assays to identify compounds which inhibit HBx activation of Pyk2 tyrosine kinase and their regulatory components and may be used to treat HBV infection and diseases and disorders associated with HBV infection.

The invention is illustrated by way of working examples which demonstrate that HBx mediates activation of a Pyk2-Src kinase signaling cascade and that activation of this signaling cascade is an essential function of HBx required to sustain HBV replication. The working examples of the present invention further demonstrate the ability of inhibitors of the Src kinase signaling cascade to inhibit HBV replication.

3.1 Definitions

As used herein, the term "target cellular gene" refers to those genes encoding members of the Src kinase family, including analogs and homologs of c-Src, Fyn, Yes and Lyn kinases and the hematopoietic-restricted kinases HcK, Fgr, LcK and Blk, and members of the Src kinase signaling pathway

including both upstream and downstream components of the Src signaling cascade, including members of the Pyk2-tyrosine kinase family. The proline rich tyrosine kinase, Pyk2, also known as cell adhesion kinase, CAK β , related adhesion focal
5 tyrosine kinase, RAFTK, and calcium-dependent protein tyrosine kinase, (CADTK), and the closely related focal adhesion kinase (FAK) comprise a family of cytoplasmic, non-receptor tyrosine kinases that can be regulated by extracellular stimuli and can activate Src-family kinases.

10 As used herein, the term "target protein" refers to those proteins which correspond to Src kinase or members of the Src kinase family or components of the Src kinase signaling pathway or proteins encoded by the HBV genome, including HBx.

15 As used herein, the terms "Src kinase" or "Src kinase family" refer to the related homologs or analogs belonging to the mammalian family of Src kinases, including, for example, the widely expressed c-Src, Fyn, Yes and Lyn kinases and the hematopoietic-restricted kinases Hck, Fgr, Lck and Blk.

20 As used herein, the terms "Src kinase signaling pathway" or "Src cascade" refer to both the upstream and downstream components of the Src signaling cascade.

25 As used herein, the term "to target" means to inhibit, block, or prevent gene expression, enzymatic activity, or interaction with other cellular or viral factors or contain a deletion or mutation in the catalytic or enzymatic portion of the target protein.

30 As used herein, the term "dominant-negative mutant" means those proteins or polypeptides which are functionally incompetent forms of the target protein and/or inhibit or modulate the enzymatic activity of the target protein or inhibit or modulate the interaction of the target protein with other cellular or viral factors.

As used herein, the term "treating or preventing HBV infection" means to inhibit the replication of the HBV virus, to inhibit HBV transmission, or to prevent HBV from establishing itself in its host, and to ameliorate or
5 alleviate the symptoms of the disease caused by HBV infection. Treating or preventing HBV infection also encompasses inhibition of viral replication in cultured cells as well as animal hosts. The treatment is considered therapeutic if there is a reduction in viral load or viral
10 pathogenesis, decrease in mortality and/or morbidity.

As used herein, the term "therapeutic agent" refers to any molecule, compound or treatment, for example and antiviral, that assists in the treatment of a viral infection or the diseases caused thereby or an agent which alleviates or assists in the treatment of a viral infection or the
15 diseases caused thereby or an agent which alleviates or assists in the treatment of disorders associated with HCC.

As used herein, the term "pharmaceutically acceptable carrier" refers to a carrier medium that does not interfere with the effectiveness of the biological activity of the
20 active ingredient, is chemically inert and is not toxic to the patient to whom it is administered.

4. BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1. Serum-starved NIH 3T3 cells were infected with AdCMV-X or -Xo viruses, c-Src or c-Fyn was
25 immunoprecipitated, the pellet washed and assayed by incubation with the substrate enolase and [γ -³²P] ATP. Products were resolved by SDS-10%-PAGE then visualized and quantitated by PhosphorImage analysis. c-Src (FIG. 1A) or c-Fyn (FIG. 1B) were immunoprecipitated 8 h p.i. from NIH 3T3
30 cell lysates expressing HBx or HBxo. Where indicated, cells were incubated with PDGF (100 ng/ml) for 5 min. (FIG. 1C) Transfected HBx activates c-Src in the cytoplasm. Chang

cells were transfected with 10 μ g pCMV-Xo (HBxo), pCMV-X (wt HBx) or pCMV-XNLS (nuclear HBx) expression plasmids, serum-starved for 24 h, c-Src immunoprecipitated and subjected to an *in vitro* trans-phosphorylation assay, then analyzed as
5 above. Similar results were obtained with NIH 3T3 cells. (FIG. 1D, FIG. 1E) c-Src (D) or c-Fyn (E) were immunoprecipitated 3 hours p.i. from Chang cells expressing HBx or HBxo. Chang cells were stimulated with 100 μ g/ml insulin (Intergen) for 10 minutes, as a positive control for Src activation (HBx mutant and HBx wildtype genes,
10 respectively).

FIGURE 2. HBx activates the Ras-MAP kinase cascade by activating the Src family of kinases. Chang cells were transfected for 18 h with 8 μ g of plasmids pCMV-Xo (HBxo) or
15 pCMV-X (wt HBx), with or without 8 μ g of plasmid pCsk or carrier DNA, then serum starved for 18 h. (FIG. 2A) c-Src was immunoprecipitated from equal amounts of cell lysates and subjected to an *in vitro* transphosphorylation assay with [γ - 32 P] ATP. (FIG. 2B) ERK2 was immunoprecipitated from equal
20 amounts of cell lysates analyzed by *in vitro* phosphorylation of myelin basic protein (MBP) using [γ - 32 P] ATP. Labeled substrate proteins were resolved by SDS-15%-PAGE, then visualized and quantitated by PhosphorImage analysis. Cells were stimulated with 100 μ g/ml of insulin (Intergen) for
25 10 min.

FIGURE 3. Transcriptional activation of AP-1 by HBx involves activation of Src kinases. Chang cells transfected with 8 μ g plasmids pCMV-X or pCMV-Xo, with or without cotransfection of pCsk for 18 h, then serum starved for 18 h.
30 (FIG. 3A) Cell extracts were prepared for AP-1 EMSA as described (Benn et al., 1996, *supra*) using a 32 P-labeled dsDNA oligonucleotide probe containing one AP-1 binding site.

Reactions were carried out using 3 μ g of nuclear extract, labeled oligonucleotide and 1 μ g of poly(dI-dC) for 30 min at 23°C. Protein-DNA complexes were resolved by electrophoresis on 4% polyacrylamide gels and visualized by PhosphorImage
5 analysis. As a positive control for AP-1 stimulation, cells were treated with 20 μ M TPA for 30 min. (FIG. 3B) Cells were transfected as above but contained in addition 3 μ g of plasmid pAP-1Luc, which encodes the luciferase reporter under the control of four AP-1 binding sites and a minimal TATA box promoter. Serum starved cells were harvested 18 h after
10 transfection and the level of expression of the luciferase reporter assayed. Results are the average of three independent experiments.

FIGURE 4. HBx strongly promotes prolonged WHV
15 replication in Chang cells. (FIG. 4A) Chang cells were transfected with 20 μ g of wt WHV or pcWHV (WHx mutant) plasmids, propagated for 11 days, intracellular core associated DNA purified through a 20% sucrose cushion, and viral DNA analyzed by Southern blot hybridization using a ³²P-
20 labeled full length genomic WHV probe. (FIG. 4B) HepG2 cells were transfected, intracellular core-associated RNA was purified 14 days post transfection, and viral DNA was analyzed as in (FIG. 4A).

FIGURE 5. *trans*-Complementation of defective WHV
25 replication by HBx. Chang cells were co-transfected with 20 μ g PCWHV (WHx mutant) and 10 μ g of either pCMV-HBxo or pCMV-HBx. Three days post-transfection, intracellular viral core particles were isolated, and viral DNA purified. Viral DNA purified from one 10 cm plate of cells was analyzed by
30 Southern blot hybridization.

FIGURE 6. Woodchuck Hepatitis B Virus (WHV) HBx protein (WHx) activates a Src-Ras signaling cascade during WHV replication in cultured cells. Chang cells were co-transfected with 20 μ g pcWHV or wtWHV with 8 μ g of either dominant-negative Ras, kinase inactive (dominant-negative) Src, or Csk plasmids. Eighteen hours post-transfection, cells were serum-starved in 0.5% CS for 20 hours, MAP kinase (ERK-2) was immunoprecipitated from equal amounts of cell lysates and pellets were subjected to an *in vitro* MBP kinase assay.

10

FIGURE 7. WHx requires activation of Src family kinase for WHV replication. Chang cells were co-transfected with 20 μ g PCWHV, wtWHV, wtWHV and RasDN (dominant-negative) (10 μ g), or wtWHV and Csk (10 μ g). Three days post-transfection viral core-associated DNA was isolated, purified, and subjected to Southern blot analysis using a full-length 32 P-labeled WHV genomic probe.

15

FIGURE 8. Cells were propagated as described (Klein, et al. 1997, EMBOJ 18: 5019-5027), transfected with 5 μ g of p Δ BS empty plasmid (vector) or pAdCMVX (HBx) expression plasmid (Klein et al., 1997, Mol. Cell. Biol. 17: 6427-6436; Klein et al., 1999, EMBOJ 18: 5019-5027; Doria et al., 1995, EMBOJ 14: 4747-4757), 2 μ g of luciferase reporter plasmid controlled by a minimal TATA-box promoter and 4 copies of an AP-1 binding site (Klein et al., 1997, Mol. Cell. Biol. 17: 6427-6436), and 5 μ g of PKM plasmid expressing a dominant-interfering form of Pyk2 or pRK5 empty plasmid (Dikic et al., 1996, Nature 383: 547-550). Cells were allowed to recover for 12 h following transfection, then serum starved for 16 h Chang cells are a human transformed hepatoblastoma line, HepG2 cells are a human differentiated hepatocytic line, and GN4 cells are a rat liver epithelial line. (A) Equal protein

20

25

30

Sab.
Bz.

amounts were assayed for luciferase activity. For analysis of low level expression of HBx, 0.4 μ g of pAdCMVX was transfected. A typical experiment is shown. (B) Cell lysates were prepared in a modified RIPA buffer (Schlaepfer et al., 1998, Mol. Cell. Biol. 18: 2571-2585), gel-electrophoresis and immunoblot analysis was performed with anti-Pyk2 or anti-Y(P)-402 Pyk2 antibodies (Biosource, Int.). Nontransfected cells were treated with 20 ng/ml TPA for 20 min to activate Pyk2 (TPA samples). (C) Equal amounts of protein lysates were immunoprecipitated with antibodies to Pyk2, an in vitro kinase assay was performed using [γ - 32 P]ATP as described (Klein, et al. 1997, EMBOJ 18: 5019-5027), and phosphorylation of associated Src, Fyn, and Pyk2 analyzed by gel-electrophoresis and autoradiography. Identification of Pyk2 and Src-Fyn proteins, which electrophoretically comigrate, was performed by immunoblot with specific antisera (not shown). (D) Fyn was immunoprecipitated and autophosphorylation activity (Fyn assay) determined by in vitro kinase assay using [γ - 32 P]ATP, gel-electrophoresis and autoradiography as described (Klein et al., 1997, Mol. Cell. Biol. 17: 6427-6436; Klein, et al. 1997, EMBOJ 18: 5019-5027). Total Fyn protein level was determined by immunoblot of an equal fraction of the immunoprecipitate.

FIGURE 9. Chang cells or HepG2 cells were transfected with expression vectors pAdCMVX (HBx), pAdCMVXo (HBx-), pPKM, or empty vectors pAdCMV or pPRK5. (A) Chang cells were treated with 50 μ M BAPTA-AM for 2 h, equal protein amounts resolved by gel-electrophoresis and immunoblotted with antibodies to Pyk2 or activated Y-402 phosphorylated Pyk2. TPA treatment was for 20 min using 20 ng/ml. (B) Chang cells treated 2 h with 50 μ M BAPTA-AM, 4 h with 3 μ M CsA, or 2 h with 0.5 mM EGTA were analyzed by immunoblot as above. (C) Chang cells transfected with vector or pAdCMVX were treated

2h with 0.5 mM or 3 mM EGTA, and Pyk2 examined by immunoblot.
(D) HepG2 cells were transfected with HBV, HBx(-) HBV genomic DNA or vector, complemented by 2 μ g plasmid pAdCMVX. Cytoplasmic HBV core particles were isolated from equal
5 numbers of cells (Klein, et al. 1997, EMBOJ 18: 5019-5027) and viral DNA replication intermediates detected by Southern blot as described (Klein, et al. 1997, EMBOJ 18: 5019-5027). The smear represents 4 kb (mature double-stranded DNA) to 2 kb (first-strand single-stranded DNA). Northern blot
10 analysis was carried out using poly(A+) RNA extracted from equal numbers of cells. (E) Southern and Northern blot analyses were performed on HepG2 cells transfected as above with or without pPKM or pPRKS. Typical experiments are shown.

15 FIGURE 10. HepG2 cells were transfected as in Figure 8 legend, allowed to recover and treated for 4 d with (A) 1 μ g/ml or 3 μ g/ml CsA, or (C) 2.5 μ M or 25 μ M BAPTA-AM. Cytoplasmic viral core particles were isolated and HBV DNA replication and mRNA levels detected by Southern and Northern
20 blot hybridization. (D) HepG2 cells were transfected with ABS or AdCMVX (HBx) plasmids and luciferase reporters containing 4 binding sites for AP-1 or CREB, linked to a TATA box promoter. Cells were treated with CsA as above and assayed for luciferase activity. (B) Endogenous polymerase
25 activity of HBV pol protein was assayed in isolated cytoplasmic core particles obtained as above, using [α - 32 P]-dNTPs in vitro as described (Nassal, 1992, J. Virol. 66: 4107-4116). Products were resolved by electrophoresis and detected by autoradiography. Typical results are shown.

30 FIGURE 11. (A) HepG2 cells were with HBV or HBx(-) HBV genomic DNA. At 24 h post-transfection cells were treated with 1 nM valinomycin (val.) or 20 nM thapsigargin (thap.)

for 4 d, cytoplasmic core particles were isolated from equal numbers of cells and viral replication examined by Southern blot hybridization. HBV transcription was analyzed by Northern blot hybridization. (B) Nontransfected HepG2 cells
5 were treated with valinomycin or thapsigargin as above, and the abundance of Pyk2 or activated Pyk2 Y-402 phosphorylation determined by immunoblot

5. DETAILED DESCRIPTION OF THE INVENTION

10 The present invention relates to therapeutic protocols and pharmaceutical compositions designed to target HBx-mediated activation of calcium-dependent tyrosine kinase, Phk2, HBx-mediated activation of Src kinase, members of the Src tyrosine kinase family and components of the Src kinase family signal transduction pathways for the treatment of HBV
15 infection and related disorders and diseases, such as HCC. The present invention relates to therapeutic protocols and pharmaceutical compositions designed to target cytosolic calcium release, regulation of calcium channels and thus, inhibit HBx-mediated activation of calcium-dependent tyrosine
20 kinase Pyk2. The invention further relates to pharmaceutical compositions for the treatment of HBV-infection targeted to HBx and its essential activities required to sustain HBV replication.

The invention is based, in part, on the Applicants' discovery that activation of Pyk2-Src kinase signaling
25 cascades plays a fundamental role in mammalian hepadnavirus replication. Applicants have demonstrated that HBx mediates activation of Pyk2, FAK, Src and MAPK signalling all occur in a calcium-dependent manner and that this activation is a critical function provided by HBx for mammalian hepadnavirus
30 replication.

The present invention encompasses a variety of protocols to inhibit HBV replication and infection, including but not

limited to: (1) protocols which target and inhibit HBx expression or inhibit the essential activities of HBx which may lead to activation of the calcium-dependent tyrosine kinase Pyk2 and the Src kinase signaling cascades; (2) 5 protocols which target and inhibit upstream effectors of the Src family of kinases, such as cytosolic calcium release, which may or may not be activated by HBx, but are required for activation of Pyk2-Src kinase signaling cascades; and (3) 10 protocols which target and inhibit Pyk2 tyrosine kinases, Src kinase family members, Src-activated enzymes and downstream effectors of Src kinases and their signal transduction pathways that are essential for viral replication.

In particular, the present invention encompasses the use of known compounds which specifically inhibit cytosolic calcium release, calcium channels, compounds which 15 specifically inhibit calcium-dependent tyrosine kinase Pyk2, and compounds which specifically inhibit the Src family of kinases and modulate activation of the Src kinase signaling cascade, including specific tyrosine kinase inhibitors, calcium channel inhibitors and Src kinase inhibitors, 20 including, but not limited to, tyrosine kinase inhibitors, drugs, organic compounds, peptides, polypeptides and nucleotides as a method of treating HBV infection and related disorders. The present invention relates to gene therapy approaches, including dominant-negative mutants, SELEX RNAs and antisense molecules targeted to Pyk2 and Src kinase 25 family members, Src-activated enzymes, downstream effectors of Src kinases and their signal transduction pathways and/or HBx.

The present invention relates to cell-based and animal model based screening assays to identify novel anti-HBV 30 agents which target HBx and its interaction and/or activation of cellular components of calcium channels, the calcium-dependent tyrosine kinase, Pyk2, and the Src kinase signaling

cascade. In addition, the present invention relates to screening assays to identify novel antiviral agents which inhibit HBx mediated activation of Src kinase and/or downstream effectors of the Src kinase signaling cascade, such as the nuclear factor, Myc. A variety of protocols and techniques may be utilized to screen for agents which interfere with and/or inhibit the interaction and/or activation of cytosolic calcium release and the Pyk2-Src kinase signaling cascade by HBx.

The present invention further encompasses pharmaceutical compositions containing the novel agents described herein. The therapeutic modalities of the invention further encompass combination therapies in which an agent which interferes with the interaction and/or activation of cytosolic calcium release and the Pyk2 and Src family of kinases by HBx, and at least one other therapeutic agent are administered either concurrently, e.g., as an admixture, separately but simultaneously or concurrently; or sequentially, including cycling therapy. Cycling therapy involves the administration of a first antiviral compound for a period of time and repeating this sequential administration, i.e., the cycle, in order to reduce the development of resistance to one of the therapies.

The novel antiviral combinations of the present invention provide a means of treatment which may not only reduce the effective dose of either drug required for antiviral activity, thereby reducing toxicity, but may also improve the absolute antiviral effect as a result of attacking the virus through multiple mechanisms. Similarly, the novel antiviral combinations provide a means for circumventing the development of viral resistance to a single therapy, thereby providing the clinician with a more efficacious treatment. Therapeutic agents to be used in combination with an agent which targets the HBx protein and

its interaction and/or activation of calcium channels and cytosolic calcium release, the Pyk2 tyrosine kinase signalling cascade and the Src kinase signaling cascade encompass a wide variety of known treatments, including
5 interferon.

**5.1 THE ROLE OF HBx MEDIATED CALCIUM DEPENDENT
ACTIVATION OF Pyk2 TYROSINE KINASE AND SRC
KINASE ACTIVATION IN HBV-INFECTION AND ITS
USE AS A TARGET FOR INTERVENTION**

10 The present invention is based, in part, on the Applicants' surprising discovery that (1) HBx requires activation of cytosolic calcium release, and calcium channels or their regulatory components to sustain HBV replication; (2) HBx acts as an intracellular cytoplasmic activator of the Pyk2 tyrosine Kinase, (3) HBx acts as an intracellular,
15 cytoplasmic activator of the Src family of nonreceptor tyrosine kinases; (4) HBx stimulates tyrosine kinase activity of the Src family kinase members, including c-Src and c-Fyn; and (5) inhibition of Src activity by the expression of a Src inhibitor, e.g., the Csk protein, results in the dramatic
20 inhibition of HBV replication. This discovery is exemplified in the in Sections 6, 7, 8 and 9 *infra*, which demonstrate that activation of Src kinase and the Src kinase signaling cascade is required to sustain HBV replication, and that inhibition of Src kinase dramatically inhibits HBV
25 replication.

Applicants have demonstrated that HBx activates Src Kinases by stimulating two related upstream tyrosine kinases known as Pyk2 and FAK. Both Pyk2 and FAK are activated by release of stored Ca²⁺ from controlled calcium channels. Applicants have discovered that HBx activity results in the
30 release of stored cytosolic Ca²⁺ from endoplasmic reticulum and/or mitochondrial channels. While not to be limited by any mechanism of activation, HBx is interacting directly or

indirectly with mitochondria, to impair voltage-dependent anion channels (VDACs), a component of the mitochondrial permeability transition pore, or Ca^{2+} channel, and is vital for maintaining mitochondrial Ca^{2+} stores, and/or endoplasmic reticulum (ER) Ca^{2+} pumps, known as SERCA pumps, which use ATP to maintain ER stores of Ca^{2+} . Thus, HBx is acting directly or through other regulatory components to partially dissipate the potential of the ER SERCA pump or the mitochondrial transitional pore resulting in cytosolic calcium release.

Applicants have demonstrated that HBx activation of Pyk2, FAK, Src and MAPK signaling all occur in a calcium-dependent manner in that treatment of cells with a calcium chelator (EGTA) or calcium channel poison (BAPTA-AM) specifically blocks HBx stimulation of Pyk2, which is essential for HBx activity. Overexpression of dominant-interfering forms of Pyk2 or FAK also block HBx transactivation activity. Further, Applicants have shown that the treatment of cells with cyclosporin A (CSA), a specific inhibitor of mitochondrial voltage-dependent anion channels, which deregulates calcium channels, also impairs HBx stimulation of HBV genomic DNA replication.

Applicants have demonstrated that HBx activation of Pyk2 promotes HBV DNA replication. The proline rich tyrosine Kinase, Pyk2, (also known as cell adhesion kinase, $CAK\beta$, related adhesion focal tyrosine kinase, RAFTK, and calcium-dependent protein tyrosine kinase (CADTK), and the closely related FAK, comprise a family of cytoplasmic, non-receptor tyrosine kinases that can be activated by extracellular stimuli and can activate Src-family kinases. Pyk2 and FAK have similarly organized domains, both having a central kinase domain flanked by extensive N-terminal and C-terminal domains. The N-terminal domains contain the major auto-phosphorylation site (the tyrosine at position 397 in FAK and

position 402 in Pyk2) which become phosphorylated upon activation of these kinases.

Applicants have demonstrated that HBx induces an increase in the tyrosine kinase activity of the Src family of kinases. The Src family of kinases, including the widely expressed Src, Fyn, Yes and Lyn kinases and the four hematopoietic-restricted kinases, Hck, Fgr, Lck and Blk (reviewed in Erpel et al. 1995 Curr. Opinion. Cell Biol. 7:176-182; Lowell et al. 1996 Genes & Devel. 10:1845-1857) are negatively regulated by phosphorylation of a carboxy-terminal residue. The Src family of kinases share common structures, which include a short amino-terminal membrane anchor, a unique domain characteristic of each individual kinase, an SH2 (Src Homology-2) domain which binds phosphotyrosine residues, an SH3 (Src Homology-3) domain which binds proline-rich sequences, a catalytic domain which has kinase activity, and a short carboxy terminal tail containing the major regulatory tyrosine residue. In addition to sharing a common overall structure, Src family kinases are regulated in a similar manner. In resting cells, the Src family of kinases are found in a repressed state in which a carboxy terminal tyrosine (y) is phosphorylated (Y-527 in c-Src (Cartwright et al. 1987 Cell 49:83-19; Kmiecik et al. 1987 Cell 49:65-74)), and naturally occurring activating mutants of Src either lacks this tyrosine or are hyperphosphorylated at this site (Iba et al. 1985 Mol. Cell. Biol. 5:1058-1066). Activated Src contains an additional phosphorylated tyrosine in the catalytic domain (Y-416 in c-Src), in which appears to be a stimulatory event *in vitro* (Cooper et al. 1993 Cell 73:1051-1054).

In the working examples described herein, the expression of HBx resulted in an increase in the ability of Src to undergo both auto- and trans-phosphorylation. The expression

of HBx also induced an increase in the tyrosine kinase activity of other Src family members, including c-Fyn.

The Applicants have further demonstrated that the expression of a Src inhibitor, i.e., the Csk protein, or
5 dominant-negative (interfering or signaling incompetent) forms of Src and Fyn proteins resulted in the inhibition of HBx mediated activation of downstream components of the Src kinase signaling cascade. Activation of Src kinase initiates a number of downstream cascades of intracellular
10 phosphorylation events. Activated Src results in activation of Ras, a prototypic member of the low-molecular weight family of protein GTPases which cycles between an inactive GDP-bound state and an active GTP-bound state. Activated Src also acts independently of the Ras signaling cascade to activate the nuclear factor Myc, among other proteins and
15 kinases (reviewed in Erpel et al., 1995 supra). The formation of active Ras-GTP complexes controls a number of downstream cellular events, including opposing cellular processes, growth and differentiation (Boguski et al. 1993 Nature 366: 643-653). Active GTP-bound Ras associates and
20 activates Raf. Activated Src has also been shown to bypass activation of Ras-GTP complexes to activate Raf in a Ras-independent manner (Stokoe & McCormick 1997 EMBOJ. 16:2384-2396). Activated Raf then phosphorylates and activates Mitogen-Activated Protein Kinase Kinase (MEK) (Dent et al. 1992 Science 257:1404-1407; Howe et al. 1992 Cell 71:335-
25 342), which in turn phosphorylates both tyrosines and threonines the extracellular-signal-regulated protein Kinases (ERKs), members of the MAP kinase (MAPK) family.

Applicants have further demonstrated that the expression a Src inhibitors, i.e., Csk protein, or dominant-negative Src
30 or Fyn proteins resulted in the inhibition of HBx activation of downstream components of Src kinase signaling cascade. Applicants have also shown that the expression of Src

dominant-negative mutants, such as Csk, inhibited the ability of HBx to stimulate activities of the nuclear factor, Myc, including stimulation of cell cycle progression by blocking HBx activation of Src kinase signaling pathways. These findings clearly establish that activation of a Src kinase signaling cascade by HBx has a critical role in the hepadnaviral life cycle.

HBx mediated activation of Src is required for HBV replication as demonstrated by way of example (Section 9 *infra*). The Applicants' work demonstrates that an essential component of the requirement of HBx viral replication in cultured cells is its ability to activate Src signaling cascades. HBx activation of a Src signaling cascade plays a critical role in transcriptional upregulation of the viral mRNAs. Inhibition of Src activity by the expression of a Src inhibitor, e.g., the CsK protein, results in the dramatic inhibition of HBV replication. These results illustrate that activation of Src family kinases has an essential role during HBV replicative life cycles.

The Applicants' discovery has implicated several targets for effective HBV anti-viral agents. HBV therapies that target the viral gene product HBx should result in a high degree of specificity and efficacy. HBV therapies that target the host cell gene products, including, but not limited to, regulatory components of cytosolic calcium release, including mitochondrial and ER calcium channels, Pyk2 kinases, the Src family of kinases, should likewise demonstrate specificity and efficacy. Although host cell gene products, Pyk2 kinases and the Src family of kinases are active in proliferating cells, such as cancer cells, or in virally infected cells. Therefore, targeting the Src family of kinases for the treatment of HBV infection should result in a high degree of efficacy, and sufficient specificity with

side effects no more toxic than chemotherapeutics currently used to treat cancer.

5.2 TREATMENT OF HBV-INFECTION USING INHIBITORS OF HBx MEDIATED SRC ACTIVATION

5 The present invention encompasses a variety of therapeutic protocols, methods and compounds to target HBx-mediated activation of cytosolic calcium release including mitochondrial and ER calcium channels, the Pyk2 signalling cascade, and the Src kinase signaling cascade for the
10 treatment of HBV. The present invention encompasses all of the compounds described in the subsections below to target HBx-mediated activation of cytosolic calcium release including mitochondrial and ER calcium channels as targets, the Pyk2 signalling cascade, and the Src kinase signaling
15 cascade with the proviso that they are not known in the art to be used to treat HBV infection, including, for example, interferon α , interferon δ , interleukin-1, interleukin-2, immune-active peptides, such as thymosin-alpha, nucleoside analogs, such as vidarabine, fialuridine, lamivuridine,
20 famciclovir, ribavirin, and corticosteroids, such as prednisone and azathioprine.

5.2.1 COMPOUNDS THAT TARGET HBx

The Applicants have demonstrated that an essential
25 activity of HBx is the activation of Src kinase signaling cascades and that this function is required for viral replication. There are a number of mechanisms by which the multi-functional HBx protein may be exerting its effects on the Src kinase signaling cascade. The present invention encompasses targeting both direct and indirect mechanisms by
30 which HBx is activating a Src kinase signaling cascade. HBx may be indirectly exerting its effects on the Src kinase signaling cascade through a variety of activities which have

FOR "FOSS" 00000000

FOR THE "905555"

been ascribed to the protein, including but not limited to transcriptional transactivating activities, binding activity to the human homolog of the UV-damage DNA repair protein involved in nucleotide excision repair, inhibitory activities
5 of a serine protease, and binding to the C7 subunit of the proteasome complex. The invention also encompasses targeting the activities of HBx which have yet to be elucidated which result in the activation of Src kinase signaling cascades. The activities of HBx are not limited to transcriptional
10 transactivation and surely other HBx associated activities remain to be discovered. Therefore, not to be limited to any theory of operation, the present invention encompasses targeting any one of the activities of HBx which are involved in activation of Src kinase signaling cascades.

For example, but not by way of limitation, compounds
15 which may be used in accordance with the present invention encompass any compound which targets HBx and inhibits its expression or interferes with its activities required for HBV replication, including but not limited to dominant-negative mutants, antisense molecules and SELEX RNAs directed to HBx.
20 The present invention further relates to nucleotides, peptides, polypeptides, fusion proteins and other compounds which further modulate HBx activities. Other examples of compounds include, but are not limited to peptide or other compounds, including small organic and inorganic molecules
25 directed to regions of the HBx protein that are required either directly or indirectly for HBx activation of Src signal cascades.

**5.2.2 COMPOUNDS THAT TARGET CYTOSOLIC CALCIUM
RELEASE AND REGULATORY COMPONENTS OF
CALCIUM CHANNELS**

30 A variety of techniques and compositions may be utilized to target to inhibit or decrease cytosolic calcium release,

including mitochondrial and ER regulation of stored Ca²⁺,
thereby inhibiting HBV replication. Such techniques and
compositions may include, but are not limited to, gene
therapy approaches, drugs, small organic molecules identified
5 to inhibit cytosolic calcium release, including mitochondrial
and ER regulation of stored Ca²⁺, and/or other upstream and
downstream effectors of cytosolic calcium release.

Sub. B3
10 In particular, compounds which may be used in accordance
with the present invention to specifically target
mitochondrial calcium channels and regulatory components
thereof. Compounds which may also be used in accordance with
the present invention include those which specifically target
endoplasmic reticulum calcium channels, SERCA Ca²⁺ pumps and
regulatory components thereof. Compounds which may be used
in accordance with the present invention include:

15 Cyclosporin A, Dihydropyridines: nifedipine (Procardia),
nimodipine (Nimotop), amlodipine (Norvasc), felodipine
(Plendil and Renedil), isradipine (DynaCirc), nifedipine
(Cardene), nisoldipine; Benzothiazepine: diltiazem
(Cardizem), Phenylalkylamine, verapamil (Calan and Isoptin),
20 Diarylaminoethylamine ethers, bepridil;
Benzimidazole-substituted tetralines, mibefradil Piperazine
, flunarizine (Sibelium); (+)-verapamil hydrochloride,
omega-Agatoxin TK, omega-Agatoxin Iva, amiloride,
Hydrochloride, nimodipine; (+)-Methoxyverapamil, ω-Agatoxin
IVA, aminohexahydrofluorene, bepridil, calcicludine,
25 calciseptine, diltiazem, flunarizine, FS2, galanin, HA 1004,
HA 1077, nifedipine, nimodipine, nitrendipine, TaiCatoxin,
protopine; cyclosporin A; BAPTA, MAPTAM, EGTA

30

5.2.3 COMPOUNDS THAT INHIBIT Pyk2 KINASE, SRC KINASE AND DOWNSTREAM EFFECTORS OF THE Pyk2-SRC KINASE SIGNALING CASCADE

30 45 60 75 90 105 120 135 150 165 180 195 210 225 240 255 270 285 300

A variety of techniques and compositions may be utilized to target Src kinase to inhibit its activity or to inhibit 5 HBx mediated activation of components of the Src kinase mediated signaling cascade, thereby inhibiting HBV replication. Such techniques and compositions may include, but are not limited to, gene therapy approaches, drugs, small organic molecules identified to inhibit Src kinase, Ras, Raf, 10 MAPK kinase, MAPK, c-Myc, cyclin-dependent kinases and/or other downstream effectors of the Src kinase signaling cascade.

115 130 145 160 175 190 205 220 235 250 265 280 295 310 325 340 355 370 385 400 415 430 445 460 475 490 505 520 535 550 565 580 595 610 625 640 655 670 685 700 715 730 745 760 775 790 805 820 835 850 865 880 895 910 925 940 955 970 985 1000

In particular, compounds which may be used in accordance with the present invention to specifically target activation 15 of Src kinase are binding proteins and competing ligands that prevent the intramolecular interaction between the carboxy-terminal phosphorylated tyrosine residue and the SH2 domain located in the amino-terminal half of the molecule and the immediately adjacent SH3 domain (Lin et al., 1993, Oncogene 8:1119-1126). In particular, compounds which may also be 20 used in accordance with the present invention include tyrosine kinase inhibitors which block the activity the Src kinase signaling cascade and therefore would block HBV replication. Examples of such tyrosine kinase inhibitors include, but are not limited to, tyrphostin-derived 25 inhibitors, which are derivatives of benzylidenemalonitrile, have been shown to have strong inhibitory activity of Src (Ramdas et al., 1995, Archives of Biochemistry and Biophysics 323:237-242), pyrazolopyrimidine PP1 (4-amino-5-(4-methylphenyl)-7-(t-butyl) pyrazolo [3,4-d] pyrimidine, a 30 selective inhibitor of the Src family of kinases (Hanke et al., 1996, J. Biol. Chem. 271:695-791) and derivatives thereof. Other examples include microbial agents, such as angelmicin B, a specific inhibitor of Src tyrosine kinase

activity, and derivatives thereof (Yokoyama et al., 1996, Leukemia Research 20:491-497), which may also be used to inhibit HBV replication.

In another embodiment of the present invention, small
5 peptides which compete with larger phosphotyrosine peptides
for binding to the Src kinase protein may be used to inhibit
the PkY2-Src kinase signaling cascade, in particular small
phosphotyrosine containing peptide ligands, 5 to 6 amino
acids, which are able to compete with larger phosphotyrosine-
10 containing peptides and protein ligands for binding to SH2
domains, thereby inhibiting the Pyk2-Src kinase signaling
cascade and blocking replication of HBV. Another embodiment
of the present invention includes small peptides which
correspond to catalytic or enzymatic domains of Pyk2 kinase,
Src kinase and would compete with the respective kinase,
15 inhibiting the activation of downstream components of the
Pyk2-Src kinase signaling cascade. Another embodiment
includes the use of larger polypeptides that inhibit Src
kinase activity including, but not limited to, Csk (carboxyl-
terminal Src kinase) which is a specific physiologic
20 inhibitor of Src kinase. Further examples of larger
polypeptides that inhibit Src kinase activity include, for
example, Src dominant-negative mutants, i.e., Srck-(Barone et
al., 1995, Nature 378:509-512) and Fyn dominant-negative
mutants (Twamley-Stein et al., 1993, Proc. Natl. Acad. Sci.
USA 90:7696-7700), also included are dominant-negative
25 mutants of downstream effectors of the Src kinase signaling
cascade, including Ras, Raf, MAPK kinase, MAPK dominant-
negative mutants and Myc dominant-negative mutants (Sawyers
et al., 1992, Cell 70:901-910).

30

**5.2.4 ANTIVIRALS TO BE USED IN COMBINATION
WITH INHIBITORS OF SRC KINASE PATHWAY**

According to the present invention, novel antiviral agents identified by the screening methods of the present invention may be used in combination with other therapeutic agents to enhance the antiviral effect achieved. Preferably a Pyk2 kinase or a Src kinase inhibitor is used in combination with another antiviral agent. Such additional antiviral agents which may be used with a Src kinase inhibitor include, but are not limited to, those which function on a different target molecule involved in viral replication, e.g., those which act on a different target molecule involved in viral transmission; those which act on a different loci of the same molecule; and those which prevent or reduce the occurrence of viral resistance. One skilled in the art would know of a wide variety of antiviral therapies which exhibit the above modes of activity.

In one embodiment of the invention, novel antiviral agents identified by the screening methods of the present invention are used in combination with known therapies to treat HBV infection, for example, IFN, interleukin-1, interleukin-2, immune-active peptides, nucleoside analogs and corticosteroids. The antiviral agents identified by the screening methods of the present invention may also be used in combination with exogenous or endogenous agents which induce IFN expression. In yet another embodiment, inhibitors of Src kinase are used in combination with agents which induce an anti-HBV immune response in order to target two different molecules required in the viral life cycle.

In order to evaluate potential therapeutic efficacy of Src kinase inhibitors in combination with the antiviral therapeutics described above, these combinations may be tested for antiviral activity according to methods known in the art.

A compound of the invention can be administered to a human patient by itself or in pharmaceutical compositions where it is mixed with suitable carriers or excipients at doses to treat or ameliorate various conditions involving HBV-infection. A therapeutically effective dose further refers to that amount of the compound sufficient to inhibit HBV infection. Therapeutically effective doses may be administered alone or as adjunctive therapy in combination with other treatments for HBV infection or associated diseases. Techniques for the formulation and administration of the compounds of the instant application may be found in "Remington's Pharmaceutical Sciences" Mack Publishing Co., Easton, PA, latest addition.

5.3 GENE THERAPY APPROACHES TO TREAT HBV INFECTION

Gene therapy approaches may also be used in accordance with the present invention to inhibit the activation of Src kinase and components of its signaling cascade. The gene therapy approaches described herein may also be applied to HBx, Src family of kinases and upstream and downstream effectors of the Src kinase signaling cascade in accordance with the present invention. Among the compounds which may disrupt the activities of HBx and its activation of the Src kinase signaling cascade are antisense, ribozyme, triple helix molecules, SELEX RNAs and dominant-negative mutants. Such molecules are designed to inhibit the expression of the target gene in HBV-infected host cells. Techniques for the production and use of antisense, ribozyme, triple helix and/or SELEX RNAs are well known to those of skill in the art and can be designed with respect to the cDNA sequence of Src kinase and components of the Src kinase signaling cascade.

5.3.1 NUCLEIC ACIDS FOR GENE THERAPY APPROACHES

Antisense RNA and DNA molecules act to directly block the translation of mRNA by hybridizing to targeted mRNA and preventing protein translation. With respect to antisense
5 DNA, oligodeoxynucleotides derived from the translation initiation site, e.g., between the -10 and +10 regions of the target gene nucleotide

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. (For a review see
10 Rossi, J., 1994, Current Biology 4:469-471). The mechanism of ribozyme action involves sequence specific hybridization of the ribozyme molecule to complementary target RNA, followed by an endonucleolytic cleavage. The composition of ribozyme molecules must include one or more sequences
15 complementary to the target gene mRNA, and must include the well known catalytic sequence responsible for mRNA cleavage. For this sequence see U.S. Patent No. 5,093,246, which is incorporated by reference in its entirety. As such, within the scope of the invention are engineered hammerhead motif ribozyme molecules that specifically and efficiently catalyze
20 endonucleolytic cleavage of RNA sequences encoding target gene proteins.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the molecule of interest for ribozyme cleavage sites which include the
25 following sequences, GUA, GUU and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the target gene containing the cleavage site may be evaluated for predicted structural features, such as secondary structure, that may render the oligonucleotide sequence unsuitable. The suitability of
30 candidate sequences may also be evaluated by testing their accessibility to hybridize with complementary oligonucleotides, using ribonuclease protection assays.

Nucleic acid molecules to be used in triple helix formation for the inhibition of transcription should be single stranded and composed of deoxynucleotides. The base composition of these oligonucleotides must be designed to promote triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of either purines or pyrimidines to be present on one strand of a duplex. Nucleotide sequences may be pyrimidine-based, which will result in TAT and CGC⁺ triplets across the three associated strands of the resulting triple helix. The pyrimidine-rich molecules provide base complementarity to a purine-rich region of a single strand of the duplex in a parallel orientation to that strand. In addition, nucleic acid molecules may be chosen that are purine-rich, for example, contain a stretch of G residues. These molecules will form a triple helix with a DNA duplex that is rich in GC pairs, in which the majority of the purine residues are located on a single strand of the targeted duplex, resulting in GGC triplets across the three strands in the triplex.

Alternatively, the potential sequences that can be targeted for triple helix formation may be increased by creating a so called "switchback" nucleic acid molecule. Switchback molecules are synthesized in an alternating 5'-3', 3'-5' manner, such that they base pair with first one strand of a duplex and then the other, eliminating the necessity for a sizeable stretch of either purines or pyrimidines to be present on one strand of a duplex.

In instances wherein the antisense, ribozyme, and/or triple helix molecules described herein are utilized to inhibit mutant gene expression, it is possible that the technique may so efficiently reduce or inhibit the transcription (triple helix) and/or translation (antisense, ribozyme) of mRNA produced by normal target gene alleles that the possibility may arise wherein the concentration of normal

FOR SEQ ID NO 1

target gene product present may be lower than is necessary for a normal phenotype. In such cases, to ensure that substantially normal levels of target gene activity are maintained, therefore, nucleic acid molecules that encode and
5 express target gene polypeptides exhibiting normal target gene activity may be introduced into cells via gene therapy methods such as those described, that do not contain sequences susceptible to whatever antisense, ribozyme, or triple helix treatments are being utilized. Alternatively,
10 in instances whereby the target gene encodes an extracellular protein, it may be preferable to coadminister normal target gene protein in order to maintain the requisite level of target gene activity.

Anti-sense RNA and DNA, ribozyme, and triple helix molecules of the invention may be prepared by any method
15 known in the art for the synthesis of DNA and RNA molecules. These include techniques for chemically synthesizing oligodeoxyribonucleotides and oligoribonucleotides well known in the art such as for example solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be
20 generated by *in vitro* and *in vivo* transcription of DNA sequences encoding the antisense RNA molecule. Such DNA sequences may be incorporated into a wide variety of vectors which incorporate suitable RNA polymerase promoters such as the T7 or SP6 polymerase promoters. Alternatively, antisense
25 cDNA constructs that synthesize antisense RNA constitutively or inducibly, depending on the promoter used, can be introduced stably into cell lines.

Various well-known modifications to the DNA molecules may be introduced as a means of increasing intracellular stability and half-life. Possible modifications include but
30 are not limited to, the addition of flanking sequences of ribo- or deoxy-nucleotides to the 5' and/or 3' ends of the molecule or the use of phosphorothioate or 2' O-methyl rather

than phosphodiesterase linkages within the oligodeoxyribonucleotide backbone.

5 Nucleic acids encoding dominant-negative mutants of the invention may be prepared by any method known in the art for the synthesis of DNA and RNA molecules. The dominant-
negative mutants of the present invention may be produced by recombinant DNA technology using techniques well known in the art. Methods which are well known to those skilled in the art can be used to construct expression vectors containing
10 the dominant-negative mutant gene product coding sequences and appropriate transcriptional and translational control signals. These methods are described in more detail herein.

5.3.2 DELIVERY OF NUCLEIC ACIDS

15 Delivery of the nucleic acid into a patient may be either direct, in which case the patient is directly exposed to the nucleic acid or nucleic acid-carrying vector, or indirect, in which case, cells are first transformed with the nucleic acid *in vitro*, then transplanted into the patient for cell replacement therapy. These two approaches are known,
20 respectively, as *in vivo* or *ex vivo* gene therapy.

In a specific embodiment, the nucleic acid is directly administered *in vivo*, where it is expressed to produce the encoded product. This can be accomplished by any of numerous methods known in the art, e.g., by constructing it as part of
25 an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, e.g., by infection using a defective or attenuated retroviral or other viral vector (see U.S. Patent No. 4,980,286), or by direct injection of naked DNA, or by use of microparticle
bombardment (e.g., a gene gun; Biolistic, Dupont), or coating
30 with lipids or cell-surface receptors or transfecting agents, encapsulation in liposomes, microparticles, or microcapsules, or by administering it in linkage to a peptide which is known

to enter the cell or nucleus, e.g., by administering it in linkage to a ligand subject to receptor-mediated endocytosis (see e.g., Wu and Wu, 1987, J. Biol. Chem. 262:4429-4432) (which can be used to target cell types specifically
5 expressing the receptors), etc. In a specific embodiment, the nucleic acid can be targeted *in vivo* for cell specific uptake and expression, by targeting a specific receptor (see, e.g., PCT Publications WO 92/06180 dated April 16, 1992 (Wu et al.); WO 92/22635 dated December 23, 1992 (Wilson et al.);
10 WO92/20316 dated November 26, 1992 (Findeis et al.); WO93/14188 dated July 22, 1993 (Clarke et al.), WO 93/20221 dated October 14, 1993 (Young)). In another embodiment, a nucleic acid-ligand complex can be formed in which the ligand comprises a fusogenic viral peptide to disrupt endosomes, allowing the nucleic acid to avoid lysosomal degradation.
15 Alternatively, the nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination (Koller & Smithies, 1989, Proc. Natl. Acad. Sci. USA 86:8932-8935; Zijlstra et al., 1989, Nature 342:435-438).
20 In a specific embodiment, a viral vector that contains the gene promoter suppressing nucleic acid is used. For example, a retroviral vector can be used (see Miller et al., 1993, Meth. Enzymol. 217:581-599). These retroviral vectors have been modified to delete retroviral sequences that are
25 not necessary for packaging of the viral genome. Retroviral vectors are maintained in infected cells by integration into genomic sites upon cell division. The nucleic acid to be used in gene therapy is cloned into the vector, which facilitates delivery of the gene into a patient. More detail about retroviral vectors can be found in Boesen et al., 1994,
30 Biotherapy 6:291-302, which describes the use of a retroviral vector to deliver the *mdr1* gene to hematopoietic stem cells in order to make the stem cells more resistant to

chemotherapy. Other references illustrating the use of retroviral vectors in gene therapy are: Clowes et al., 1994, J. Clin. Invest. 93:644-651; Kiem et al., 1994, Blood 83:1467-1473; Salmons and Gunzberg, 1993, Human Gene Therapy 4:129-141; and Grossman and Wilson, 1993, Curr. Opin. in Genetics and Devel. 3:110-114.

In yet another specific embodiment, attenuated viruses, such as hepadnaviruses, which have the same tropism as HBV, may be engineered and used for gene therapy in accordance with the present invention. Hepadnaviruses are particularly attractive for use in gene therapy in accordance with the present invention as these viruses will deliver the therapeutic exactly to those cells infected with HBV. Hepadnaviral vectors would be particularly effective for the delivery of nucleic acids targeting components of the Src kinase signaling cascade, thereby avoiding unnecessarily knocking out expression of host genes.

Adenoviruses are other viral vectors that can be used in gene therapy. Adenoviruses are especially attractive vehicles for delivering genes to liver and respiratory epithelia. Adenoviruses naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are liver, the central nervous system, endothelial cells, and muscle. Adenoviruses have the advantage of being capable of infecting non-dividing cells. Kozarsky and Wilson, 1993, Current Opinion in Genetics and Development 3:499-503 present a review of adenovirus-based gene therapy. Bout et al., 1994, Human Gene Therapy 5:3-10 demonstrated the use of adenovirus vectors to transfer genes to the respiratory epithelia of rhesus monkeys. Other instances of the use of adenoviruses in gene therapy can be found in Rosenfeld et al., 1991, Science 252:431-434; Rosenfeld et al., 1992, Cell 68:143-155; and Mastrangeli et al., 1993, J. Clin. Invest, 91:225-234.

Adeno-associated virus (AAV) has also been proposed for use in gene therapy (Walsh et al., 1993, Proc. Soc. Exp. Biol. Med. 204:289-300. Herpes viruses are other viruses that can also be used.

5 Another approach to gene therapy, for use in the cell replacement therapy of the invention, involves transferring a gene to cells in tissue culture by such methods as electroporation, lipofection, calcium phosphate mediated transfection, or viral infection. Usually, the method of transfer includes the transfer of a selectable marker to the
10 cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene. Those cells are then delivered to a patient.

In this embodiment, the nucleic acid is introduced into
15 a cell prior to administration *in vivo* of the resulting recombinant cell. Such introduction can be carried out by any method known in the art, including, but not limited to, transfection, electroporation, microinjection, infection with a viral vector containing the nucleic acid sequences, cell
20 fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion, etc. Numerous techniques are known in the art for the introduction of foreign genes into cells (see e.g., Loeffler & Behr, 1993, Meth. Enzymol. 217:599-618; Cohen et al., 1993, Meth. Enzymol. 217:618-644; Cline, 1985, Pharmac. Ther. 29:69-92) and may be used in
25 accordance with the present invention, provided that the necessary developmental and physiological functions of the recipient cells are not disrupted. The technique should provide for the stable transfer of the nucleic acid to the cell, so that the nucleic acid is expressible by the cell and
30 preferably heritable and expressible by its cell progeny.

The resulting recombinant cells can be delivered to a patient by various methods known in the art. In a preferred

embodiment, epithelial cells are injected, e.g., subcutaneously. In another embodiment, recombinant skin cells (e.g., keratinocytes) may be applied as a skin graft onto the patient. Recombinant blood cells (e.g.,
5 hematopoietic stem or progenitor cells) are preferably administered intravenously. The amount of cells envisioned for use depends on the desired effect, patient state, etc., and can be determined by one skilled in the art.

In an embodiment in which recombinant cells are used in gene therapy, nucleotides which encode a gene or promoter
10 suppressor are introduced into the cells such that it is expressible by the cells or their progeny, and the recombinant cells are then administered *in vivo* for therapeutic effect. In a specific embodiment, stem or progenitor cells are used. Any stem and/or progenitor cells
15 which can be isolated and maintained *in vitro* can potentially be used in accordance with this embodiment of the present invention.

5.4 PHARMACEUTICAL FORMULATIONS AND 20 METHODS OF ADMINISTRATION

The present invention encompasses the use of known agents which block HBx activation of Src kinase signaling cascade and novel antiviral agents identified by the screening methods of the invention in pharmaceutical compositions and therapeutic modalities for the treatment of
25 HBV infection, and the disorders and diseases associated with HBV infection, including HCC. In one embodiment of the present invention, the novel antiviral agents identified by the screening assays of the present invention may be used in combination with other known antiviral agents to treat viral
30 infections.

5.4.1 ROUTES OF ADMINISTRATION

Suitable routes of administration may, for example, include oral, rectal, transmucosal, or intestinal administration; parenteral delivery, including intramuscular, 5 subcutaneous, intramedullary injections, as well as intrathecal, direct intraventricular, intravenous, intraperitoneal, intranasal, or intraocular injections, and optionally in a depot or sustained release formulation.

Furthermore, one may administer the agent of the present invention in a targeted drug delivery system, for example in 10 a liposome targeted to the liver. The liposomes will be targeted to and taken up selectively by liver cells.

5.4.2 COMPOSITION/FORMULATION

The pharmaceutical compositions of the present invention 15 may be manufactured in a manner that is itself known, e.g., by means of conventional mixing, dissolving, dragee-making, levitating, emulsifying, encapsulating, entrapping, or lyophilizing processes.

Pharmaceutical compositions for use in accordance with 20 the present invention thus may be formulated in conventional manner using one or more physiologically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Proper formulation is dependent upon the route of administration chosen. 25

For injection, the agents of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers, such as Hank's solution, Ringer's solution, or physiological saline buffer. For 30 transmucosal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are usually known in the art.

For oral administration, the compounds can be formulated readily by combining the active compounds with pharmaceutically acceptable carriers well known to those in the art.

5 Such carriers enable the compounds of the invention to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for oral ingestion by a patient to be treated. Pharmaceutical preparations for oral use can be obtained solid excipient, optionally grinding a resulting mixture, and processing the
10 mixture of granules, after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are, in particular, fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat
15 starch, rice starch, potato starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, sodium carboxymethylcellulose, and/or polyvinylpyrrolidone (PVP). If desired, disintegrating agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, or alginic acid or
20 a salt thereof such as sodium alginate.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used, which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, and/or
25 titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

Pharmaceutical preparations which can be used orally
30 include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules can contain the

active ingredients in admixture with filler such as lactose,
binders such as starches, and/or lubricants such as talc or
magnesium stearate and, optionally, stabilizers. In soft
capsules, the active compounds may be dissolved or suspended
5 in suitable liquids, such as fatty oils, liquid paraffin, or
liquid polyethylene glycols. In addition, stabilizers may be
added. All formulations for oral administration should be in
dosages suitable for such administration.

For buccal administration, the compositions may take the
10 form of tablets or lozenges formulated in conventional
manner.

For administration by inhalation, the compounds for use
according to the present invention are conveniently delivered
in the form of an aerosol spray presentation from pressurized
packs or a nebulizer, with the use of a suitable propellant,
15 e.g., dichlorodifluoromethane, trichlorofluoromethane,
dichlorotetrafluoroethane, carbon dioxide or other suitable
gas. In the case of a pressurized aerosol the dosage unit
may be determined by providing a valve to deliver a metered
amount. Capsules and cartridges of e.g. gelatin for use in
20 an inhaler or insufflator may be formulated containing a
powder mix of the compound and a suitable powder base such as
lactose or starch.

The compounds may be formulated for parenteral
administration by injection, e.g., by bolus injection or
continuous infusion. Formulations for injection may be
25 presented in unit dosage form, e.g., in ampoules or in multi-
dose containers, with an added preservative. The
compositions may take such forms as suspensions, solutions or
emulsions in oily or aqueous vehicles, and may contain
formulatory agents such as suspending, stabilizing and/or
30 dispersing agents.

Pharmaceutical formulations for parenteral
administration include aqueous solutions of the active

compounds in water-soluble form. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Optionally, the suspension may also contain suitable stabilizers or agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, e.g., sterile pyrogen-free water, before use.

The compounds may also be formulated in rectal compositions such as suppositories or retention enemas, e.g., containing conventional suppository bases such as cocoa butter or other glycerides.

In addition to the formulations described previously, the compounds may also be formulated as a depot preparation. Such long acting formulations may be administered by implantation (for example subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compounds may be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

Liposomes and emulsions are well known examples of delivery vehicles or carriers for hydrophobic drugs. Certain organic solvents such as dimethylsulfoxide also may be employed, although usually at the cost of greater toxicity. Additionally, the compounds may be delivered using a

sustained-release system, such as semipermeable matrices of solid hydrophobic polymers containing the therapeutic agent. Various of sustained-release materials have been established and are well known by those skilled in the art. Sustained-
5 release capsules may, depending on their chemical nature, release the compounds for a few weeks up to over 100 days. Depending on the chemical nature and the biological stability of the therapeutic reagent, additional strategies for protein stabilization may be employed.

10 The pharmaceutical compositions also may comprise suitable solid or gel phase carriers or excipients. Examples of such carriers or excipients include but are not limited to calcium carbonate, calcium phosphate, various sugars, starches, cellulose derivatives, gelatin, and polymers such as polyethylene glycols.

15 Many of the compounds of the invention identified as inhibitors of the Src kinases may be provided as salts with pharmaceutically compatible counterions. Pharmaceutically compatible salts may be formed with many acids, including but not limited to hydrochloric, sulfuric, acetic, lactic,
20 tartaric, malic, succinic, etc.; or bases. Salts tend to be more soluble in aqueous or other protonic solvents that are the corresponding free base forms. Examples of pharmaceutically acceptable salts, carriers or excipients are well known to those skilled in the art and can be found, for example, in Remington's Pharmaceutical Sciences, 18th
25 Edition, A.R. Gennaro, Ed., Mack Publishing Co., Easton, PA, 1990. Such salts include, but are not limited to, sodium, potassium, lithium, calcium, magnesium, iron, zinc, hydrochloride, hydrobromide, hydroiodide, acetate, citrate, tartrate, malate sales, and the like.

30

5.4.3 EFFECTIVE DOSAGE

Pharmaceutical compositions suitable for use in the present invention include compositions wherein the active ingredients are contained in an effective amount to achieve
5 their intended purpose. More specifically, a therapeutically effective amount means an amount effective to prevent development of or to alleviate the existing symptoms of the subject being treated. Determination of the effective amounts is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided
10 herein.

For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. Such information can be used to more accurately determine useful doses in humans.

15 A therapeutically effective dose refers to that amount of the compound that results in a reduction in the intensity of the infection or in amelioration of symptoms or a prolongation of survival in a patient. Toxicity and therapeutic efficacy of such compounds can be determined by
20 standard pharmaceutical, pharmacological, and toxicological procedures in cell cultures or experimental animals, e.g., for determining the LD₅₀ (the dose lethal to 50% of the population) and the ED₅₀ (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and
25 therapeutic effects is the therapeutic index and it can be expressed as the ratio between LD₅₀ and ED₅₀. Compounds which exhibit high therapeutic indices are preferred. The data obtained from cell culture assays or animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of
30 circulating concentrations that include the ED₅₀ with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of

administration utilized. The exact formulation, route of administration and dosage can be chosen by the individual physician in view of the patient's condition. (See e.g. Fingl et al., 1975, in "The Pharmacological Basis of Therapeutics", Ch. 1 p. 1).

Sub. B. 10 Dosage amount and interval may be adjusted individually to provide plasma levels of the active moiety which are sufficient to maintain the desired modulating effects, or minimal effective concentration (MEC). The MEC will vary for each compound but can be estimated from in vitro data; e.g., the concentration necessary to achieve a 50-90% inhibition of HCV infection using the assays described herein. Dosages necessary to achieve the MEC will depend on individual characteristics and route of administration. However, HPLC assays, bioassays or immunoassays can be used to determine plasma concentrations.

20 Dosage intervals can also be determined using the MEC value. Compounds should be administered using a regimen which maintains plasma levels above the MEC for 10-90% of the time, preferably between 30-90% and most preferably between 50-90%.

In cases of local administration or selective uptake, the effective local concentration of the drug may not be related to plasma concentration.

25 The amount of composition administered will, of course, be dependent on the subject being treated, on the subject's weight, the severity of the affliction, the manner of administration and the judgment of the prescribing physician.

30 In immunization procedures, the amount of immunogen to be used and the immunization schedule will be determined by a physician skilled in the art and will be administered by reference to the immune response and antibody titers of the subject.

5.4.4. PACKAGING

The compositions may, if desired, be presented in a pack or dispenser device which may contain one or more unit dosage forms containing the active ingredient. The pack may for
5 example comprise metal or plastic foil, such as a blister pack. The pack or dispenser device may be accompanied by instructions for administration. Compositions comprising a compound of the invention formulated in a compatible pharmaceutical carrier may also be prepared, placed in an
10 appropriate container, and labelled for treatment of an indicated condition.

5.5 SCREENING ASSAYS TO IDENTIFY NOVEL HBV-ANTIVIRAL AGENTS

At least two different assay systems, described in the
15 subsections below, can be designed and used to identify compounds or compositions that modulate HBx-mediated activation of Src kinase signaling cascades and thereby inhibit HBV replication.

The systems described below may be formulated into kits.
20 To this end, cells expressing HBx and components of the Src kinase signaling cascade, or cells expressing components of the Src kinase signaling cascade which are capable of sustaining HBV replication, or cell lysates thereof can be packaged in a variety of containers, e.g., vials, tubes, microtitre well plates, bottles, and the like. Other
25 reagents can be included in separate containers and provided with the kit; e.g., positive control samples, negative control samples, buffers, cell culture media, etc.

30

5.5.1 CELL-BASED ASSAYS

In accordance with the invention, a cell-based assay system can be used to screen for compounds that target HBx and/or HBx-mediated activation of Src kinase signaling cascades. To this end, cells that endogenously express components of the Src kinase signaling cascade, including for example, Src kinase, Ras, Raf kinase, MAPK kinase, MAPK, JNK, Myc and cyclin-dependent kinases can be engineered to express HBx under the control of an inducible promoter. Alternatively, cell lines may be engineered to express a component or components of the Src signaling cascade under the control of an inducible promoter. The cloning and characterization for each component has been described: (See, for example, Erpel et al., 1995, J. Biol. Chem. 271:16807-16812; Boquski et al., 1993, Nature 366:643-653; Bruder et al., 1992, Genes Dev. 6:545-556; Derijord et al., 1994 Cell 76:1025-1037; Miden et al, 1994 Science 266:1719-1723) each of which is incorporated by reference herein in its entirety.

The invention further encompasses cell lines, expressing both HBx and Src, either inducibly or constitutively, which results in cell cultures which may be maintained for both the short term and long term support of HBV replication. The cell lines of the present invention support HBV replication and may be maintained for long periods of time, have utility for the study of HBV replication, i.e., to identify additional cellular components required to support HBV replication, in addition to identifying potential antiviral agents for the treatment of HBV infection.

Alternately, cell lines which co-express HBx and Src kinase and components of the Src kinase signaling cascade may be genetically engineered to assay for inhibitors of HBx activation of Src. This can be engineered in cell in the

absence of HBV replication or in cell lines which support the HBV life cycle as a means of (1) identifying additional factors required to support the HBV life cycle, and (2) as a system to screen test compounds, for their ability to
5 interfere with HBx activation and/ or interaction with the Src kinase, and (3) as a system to screen test compounds for their ability to inhibit Src kinase activity and therefore inhibit HBV replication.

The present invention encompasses the expression of Src
10 kinase and components of the Src kinase signaling cascade in cell lines. In a preferred embodiment of the invention, HBx and Src kinase and components of the Src kinase signaling cascade are co-expressed in cell lines, such as human hepatoma cell lines, HepG2 and Huh7.

Methods which are well known to those skilled in the art
15 can be used to construct expression vectors containing HBx or Src kinase gene product coding sequences and appropriate transcriptional and translational control signals. In accordance with the present invention, nucleotides encoding HBx and/or Src kinase and components of the Src kinase
20 signaling cascade may be expressed under the control of an inducible promoter. These methods include, for example, in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. See, for example, the techniques described in Sambrook et al., 1989, supra, and Ausubel et al., 1989, supra. Alternatively, RNA capable of
25 encoding Src kinase gene product sequences may be chemically synthesized using, for example, synthesizers. See, for example, the techniques described in "Oligonucleotide Synthesis", 1984, Gait, M.J. ed., IRL Press, Oxford, which is incorporated by reference herein in its entirety.

30 In a preferred embodiment of the present invention, human Src kinase and HBx are co-expressed in hepatic cell lines, such as human hepatoma cell lines, HepG2 and Huh7, and

Chang liver cells. A number of selection systems well known to those skilled in the art may be used to successfully engineer cell lines which express the Src kinase and HBx gene products. Appropriate cell lines or host systems can be
5 chosen to ensure the correct modification of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such mammalian host cells include but are not limited to CHO, VERO, BHK, HeLa, COS, MDCK, 293,
10 3T3 and WI38 cell lines.

In a preferred embodiment of the in vitro screening methods of the present invention, test methods rely on measurements of Src kinase activity or by measuring the activity of downstream effectors of Src kinase, such as MAP
15 kinase and Myc. The assays of the present invention may include in vitro kinase assays which measure the effects of test compounds on individual components of the Src kinase signaling pathway. For example, but not by way of limitation, these assays may involve measuring the effects of
20 a test compound on a reaction mixture containing a cell lysate prepared from HBV infected cells treated with the test compound. Src kinase may be immunoprecipitated from the cell lysate and c-Src autophosphorylation or enolase transphosphorylation activity of the immunoprecipitated
25 kinase may be determined. MAP kinase or another component of the Src Kinase pathway may be immunoprecipitated from the cell lysate and its kinase activity measured using a known substrate of the kinase, i.e. myelin basic protein or transcription factors AFT-2, CHOP, HSP27 and Max. The activity of the MAP kinase will be determined as a
30 measurement of the phosphorylated state of the substrate. In order to test a compound for inhibitory activity, the reaction mixture is prepared in the presence and absence of

the test compound. Control reaction mixtures are incubated without the test compound or with a placebo. The lack of phosphorylation of the substrate indicates that the MAP kinase is inhibited indicating that the Src kinase signaling pathway is blocked.

In yet another embodiment of the invention, the ability of a test agent to inhibit HBx activation of Src's induction of downstream effectors may be measured. For example, activation of Src kinase signaling cascade leads to enhanced expression of Myc proteins. Therefore, activation of Myc promoter elements may be used to determine the potential anti-HBV activity of the test agent. Constructs encoding the Myc promoter region linked to any of a variety of different reporter genes may be introduced into cells expressing the Src kinase and/or components of the Src kinase signaling cascade. Such reporter genes may include but is not limited to chloramphenicol acetyltransferase (CAT), luciferase, GUS, growth hormone, or placental alkaline phosphatase (SEAP). Following exposure of the cells to the test compound, the level of reporter gene expression may be quantitated to determine the test compound's ability to regulate receptor activity. Alkaline phosphatase assays are particularly useful in the practice of the invention as the enzyme is secreted from the cell. Therefore, tissue culture supernatant may be assayed for secreted alkaline phosphatase. In addition, alkaline phosphatase activity may be measured by calorimetric, bioluminescent or chemiluminescent assays such as those described in Bronstein, I. et al. (1994, Biotechniques 17: 172-177). Such assays provide a simple, sensitive easily automatable detection system for pharmaceutical screening.

In yet another embodiment of the invention, measurements of the tyrosine phosphorylated state of Src may be used to measure the effects of test compounds on the Src kinase

signaling pathway. In resting cells, the Src family of kinases are found in a repressed state in which a carboxy terminal tyrosine (Y) is phosphorylated (Y-527 in c-Src). Active Src is hypophosphorylated at this residue, therefore, phosphorylation of this residue of c-Src in response to a test agent may indicate the ability of that test agent to inhibit c-Src activation. Activated Src contains an additional phosphorylated tyrosine in the catalytic domain (Y-416 in c-Src), which appears to be a stimulatory event *in vitro* (Cooper et al., 1993, Cell 73:1051-1054). Therefore, dephosphorylation of this residue of c-Src in response to a test agent may indicate the ability of that test agent to inhibit c-Src activation.

In a particular embodiment, the target substrate can be prepared for immobilization using recombinant DNA techniques routinely used in the art. The target substrate may be for example, a region of Src containing the carboxy terminal tyrosine residue or a region of MAPK containing the tyrosine residue. For example, the target gene coding region can be fused to a glutathione-S-transferase (GST) gene using a fusion vector, such as pGEX-5X-1, in such a manner that its conformation is maintained in the resulting fusion protein. In such an assay, e.g., the target gene fusion protein can be anchored to glutathione-agarose beads. The activated or infected cell lysate can be added in the presence or absence of the test compound in a manner that allows the kinase reaction to occur in the presence of ³²P-ATP. At the end of the reaction period, unbound material can be washed away, and the substrate assayed for its phosphorylated state. The interaction between the target gene protein and the Src kinase pathway component can be detected by measuring the amount of radioactivity that remains associated with the glutathione-agarose beads. A successful inhibition of the

interaction by the test compound will result in a decrease in measured radioactivity.

In another embodiment of the cell-based assays of the present invention, activation of Src kinase signaling cascades mediated by HBx may be measured by a viability assay to positively test for effective compounds. For example, a test compound may be applied to cells expressing HBx and Src kinase signaling components. Then an agent which induces cell death in response to activated Src kinase, such as tumor necrosis factor (TNF), is applied to cells. If the test compound is ineffective in inhibiting HBx mediated activation of Src kinase, the cells die. Compounds effective in inhibiting activation, result in cell viability. Such an assay system provides a quick and easy read-out to determine the effectiveness of a test compound to inhibit HBx mediated activation of Src kinase.

Alternatively, activation of Src kinase signaling pathways mediated by HBx may be measured by the secretion of mature HBV viral particles into the medium of growing Chang cells. For example, Chang liver cells may be stably transformed with an HBV or WHV pregenome, or with a head-to-tail dimer of either HBV or WHV genomes. The integrated virus will produce and secrete HBV/WHV particles into the medium. As demonstrated by the Applicants, the secretion of viral particles is strongly enhanced by HBx protein activation of Src kinases. If the test compound is effective in inhibiting HBx activation of Src, it will result in reduced secretion of HBV/WHV particles into the medium. The level of particle secreted into the medium can be assayed using commercial ELISA kits to detect the presence of HBV/WHV HBcAg and HBsAg.

Alternatively, the activation of Src kinase signaling pathways mediated by HBx may be measured in fission yeast, *Schizosaccharomyces pombe*. Src kinase have not been found in

single cell lower eukaryotes such as yeast, and their expression induces cell death of the fission yeast (Erpel, T., Superti-Furga, G. & Courtneidge, S.A., 1995, EMBO J. 14:963-975). Studies have shown that yeast cell viability is maintained only by inhibition of Src activation, for instance, by coexpression with the Src inhibitor Csk (Koegl, M., Courtneidge, S.A. & Superti-Furga, G. 1996, Oncogene 11:2317-2329). Therefore, a viability assay is based on the fact that activation of Src kinase by HBx protein in yeast will result in cell death, whereas the inhibition of Src kinase will permit cells to grow and reproduce. *S. pombe* can be transformed with the c-Src gene and the HBx gene under the control of regulated promoters. A variety of regulated promoters can be chosen, such as the thiamine repressible *nmt 1* promoter. Removal of thiamine from the medium will result in induction of both HBx and c-Src proteins, and subsequent killing of cells. If a test compound is effective in inhibiting HBx activation of Src, it will block the growth arrest.

20 5.5.2 ANIMAL MODEL SCREENING ASSAYS

The present invention relates to animal model screening assays to identify compounds effective to inhibit HBV replication. In accordance with these animal model screening assays, the present invention encompasses the expression of Src kinase and components of the Src kinase signaling cascade with or without the co-expression of HBx.

Animals of any species, including, but not limited to, woodchucks, mice, rats, rabbits, guinea pigs, pigs, micro-pigs, goats, and non-human primates, e.g., baboons, monkeys, and chimpanzees may be used to generate HBx and Src kinase transgenic animals.

Any technique known in the art may be used to introduce the HBx and Src kinase gene transgene into animals to produce

the founder lines of transgenic animals. Such techniques include, but are not limited to pronuclear microinjection (Hoppe, P.C. and Wagner, T.E., 1989, U.S. Pat. No. 4,873,191); retrovirus mediated gene transfer into germ lines
5 (Van der Putten et al., 1985, Proc. Natl. Acad. Sci., USA 82:6148-6152); gene targeting in embryonic stem cells (Thompson et al., 1989, Cell 56:313-321); electroporation of embryos (Lo, 1983, Mol Cell. Biol. 3:1803-1814); and sperm-mediated gene transfer (Lavitrano et al., 1989, Cell 57:717-723); etc. For a review of such techniques, see Gordon,
10 1989, Transgenic Animals, Intl. Rev. Cytol. 115:171-229, which is incorporated by reference herein in its entirety.

In preferred embodiment of the invention, Src kinase is expressed alone in transgenic Src knock-out mice and a HBV pseudovirus is used to infect the animals. For example, a
15 HBV pseudovirus which contains the HBV virus and an envelope protein from a virus with a natural tropism for murine cells, such as the murine leukemia virus (MLV), is used to bypass internalization of the HBV virus by the murine cells. These murine cells can then support the life cycle of the
20 internalized HBV virus, because they express human Src kinase.

In one embodiment of the animal model screen of the present invention, test compounds may be administered to transgenic animals and their ability to inhibit HBV
25 replication directly measured. For example, but not by way of limitation, these assays may involve measuring the effects of test compounds on viral replication in host cells and transgenic animals. HBV replication may be measured by a number of criteria, including measuring viral DNA. Intracellular core-associated DNA may be purified and
30 analyzed by southern blot hybridization to measure synthesis of viral DNA replicative intermediates. The synthesis of viral DNA replicative intermediates in the presence of test

compounds may be compared to levels of viral DNA replicative intermediates in the absence of the test compounds. Levels of HBV replication in the presence or absence of test compounds may also be determined by measuring levels of
5 extracellular virus which is released or intracellular viral transcripts and/or viral proteins, including the surface antigen and the polymerase protein, using standard molecular biological techniques and methods known to those of ordinary skill in the art.

10 In yet another embodiment of the animal model screens of the present invention, the effect of test compounds to inhibit HBV replication may be measured indirectly. For example, transgenic mice may be engineered which express (1) the HBx gene product under the control of an inducible promoter, and (2) readout vector which is responsive to Src
15 activation. The readout vector may comprise a reporter gene under the control of a Myc promoter. Such reporter constructs are described in Section 5.5.1 *infra*. In this assay system, expression of the HBx gene product is induced and the test compound is administered to the mice. The
20 ability of the test compound to inhibit HBx mediated activation of Src kinase and HBV replication is assayed by measuring the reporter gene. Such reporter genes may include but are not limited to chloramphenicol acetyltransferase (CAT), luciferase, GUS, growth hormone, or placental alkaline phosphatase (SEAP).
25 Following exposure of the animal to the test compound, the level of reporter gene expression may be quantitated from the blood or tissue sample to determine the test compound's ability to regulate receptor activity. Alkaline phosphatase assays are particularly useful in the practice of the invention as the enzyme is secreted from the
30 cell. Therefore, tissue culture supernatant may be assayed for secreted alkaline phosphatase. In addition, alkaline phosphatase activity may be measured by calorimetric,

095906-0414

bioluminescent or chemiluminescent assays such as those described in Bronstein, I. et al. (1994, Biotechniques 17: 172-177). Such assays provide a simple, sensitive easily detection system for pharmaceutical screening.

5

6. **EXAMPLE: HBx IS AN INTRACELLULAR ACTIVATOR OF SRC KINASE AND DOWNSTREAM COMPONENTS OF THE SRC SIGNALING CASCADE**

The following studies demonstrate that HBx activates the Src family of nonreceptor tyrosine kinases. These studies also demonstrate that activation of Src fundamentally mediates HBx activation of Ras signaling and cell cycle progression.

10

6.1 MATERIALS AND METHODS

Cell Culture

15

Cell lines used in this study were obtained from the American Tissue Type Culture Collection. Chang liver, NIH 3T3, and 293 cells were propagated in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% bovine calf serum (CS) and 50 mg/ml gentamycin sulfate. HepG2 cells were propagated in DMEM supplemented with 10% fetal CS. Serum starvation of Chang cells was carried out in 0.05% CS for 24-48 hours, and for NIH 3T3 cells, in 0.5% CS for 16-20 hours.

20

25 Transfections and Adenovirus Infections

Chang cells were transfected by the calcium phosphate precipitation technique (Wigler et al., 1979 Proc. Natl. Acad. Sci. 76:1373-1376). 6×10^5 - 7.5×10^5 Chang cells per 10 cm plate were transfected for 7-10 hours (20 μ g DNA total), after which the media was aspirated and replaced with fresh 10% CS/DMEM (Doria et al., 1995 EMBO J 14:4747-4759). Eighteen hours after transfection, Chang cells were serum starved either in 0.05% CS for 16-20 hours followed by

30

1238500

addition of 2% serum for 2 hours to allow protein synthesis,
or serum starved in 0.5% CS for 24 hours and then harvested.
NIH 3T3 cells were transfected by the calcium phosphate
technique overnight, serum starved in 0.5% CS for 18 hours
5 and infected with the recombinant adenoviruses (described
below).

NIH 3T3 cells were plated at 1×10^6 cells/10cm plate
and serum starved the next day in 0.5% CS for 16-18 hours.
Chang cells were also plated at 1×10^6 cells/10cm plate and
serum starved in 0.05% CS for 24-48 hours. Cells were
10 infected with AdCMV-X or AdCMV-Xo viruses at 25 pfu for 1
hour in 1 ml of PBS/2% CS, after which 10 ml of 10% CS/DMEM
was added and the cells harvested at the indicated times.

The replication-defective recombinant adenoviruses
express, under the control of the CMV promoter, either wild
15 type HBx (AdCMV-X) or a mutated HBx mRNA in which all the
AUGs have been mutated (AdCMV-Xo). The construction of the
recombinant adenoviruses, AdCMV-X and AdCMV-Xo, have been
previously described (Doria et al., 1995, supra). Viruses
were propagated and titered in 293 cells before use.

20
Antibodies and Plasmids

Ras-specific monoclonal antibody Y13-259. (Santa Cruz
Biochemicals, Inc.). Rabbit anti-Shc and anti-Sos serum.
(Santa Cruz Biochemicals, Inc.). Rabbit anti-Grb2,
25 anti-ERK2, anti-Csk, anti-JNK and anti-N-Myc were purchased
from Santa Cruz. Anti-Src (M327) antibodies were purchased
from Oncogene Science. Anti-Fyn antibodies (Santa Cruz) were
a gift of D. Littman (NYU). Anti-phosphotyrosine (4G10; UBI)
and rabbit anti-p34cdc2 serum and Rabbit anti-RPTP α serum.
(Upstate Biotech Inc.).

30 Plasmid pHSos expresses human hSos under the CMV
promoter plasmid pCaCSK expresses the CSK gene under the
control of the CMV promoter. Plasmid pSRC expresses c-Src

FOR "SECRET" 60

Sub. 25

under the control of the CMV promoter. Plasmids pCMVSK- and pCMVFK- express the kinase-inactive (dominant-negative) forms of Src (SK-) and Fyn (FK-) under the control of the SV-40 LTR, respectively (Twamley-Stein et al., 1993 Proc. Natl. Acad. Sci. 90:696-700). For the present studies, the cDNAs for the kinase-inactive forms of Src and Fyn were subcloned into pCDNA1Amp (Invitrogen) for expression under the control of the CMV promoter. To create pCMVSK- (containing the dominant-negative Src under the control of the CMV promoter), a BamH1/Sal1 fragment containing the entire kinase-inactive Src was ligated into BamH1/Xho1 sites in pCDNA1Amp. To create pCMVFK- (containing the dominant-negative Fyn under the control of the CMV promoter), a BamH1 fragment containing the entire kinase-inactive Fyn was ligated into the BamH1 site of pCDNA1Amp. Plasmids wtWHV expresses the wild-type WHV genome. Plasmids pAUGWHV and pcWHV expresses WHV genome with mutations that ablate WHx expression (Seeger et al. 1991 J. Virol. 65:1673-1679). Plasmid pAUGWHV contains a mutation in the first AUG of the X open reading frame (ORF) plasmid pcWHV contains a stop codon between the second and third AUG in the X ORF (Zoulim et al., 1994, J. Virol. 68:2026-2030).

pCMV-X expresses wild-type HBx under the control of the CMV promoter. pCMV-Xo contains a mutant HBx gene in which all of the AUGs in the ORF have been mutated to GUG and therefore a functional HBx protein cannot be translated from the mRNA (Doria et al., 1995, supra). HBx-NLS contains a functional nuclear localization signal and exclusively relocates HBx protein to the nucleus, whereas HBx-SLN expresses a mutated NLS signal and is a cytoplasmic protein (Doria et al., 1995, supra). The expression vectors pCMV-HBx-NLS and pCMV-HBx-SLN were constructed by replacing the adenovirus major late promoter with the CMV promoter in the vectors pAd-HBx-NLS and pAd-HBx-SLN by ligating an

EcoRI/SalI fragment from pCMV3C containing the CMV promoter and the adenovirus tripartite leader into the corresponding EcoRI/SalI sites in pAd-HBx-NLS/SLN.

5 Immunoprecipitation and Western Immunoblotting

Cells were lysed in 1% Triton, 20 mM Hepes pH 7.4, 150 mM NaCl, 10% Glycerol, 1 mM Na₃VO₄, 50 mM NaF, 1 mM PMSF, 10 μg/ml aprotinin, and 10 μg/ml leupeptin. Equal amounts of protein were immunoprecipitated on ice for 2 hours and the immune complexes were collected by incubation with protein A-agarose (Santa Cruz) for 60 minutes at 4°C.

Immunoprecipitates were washed three times with HNTG (20 mM Hepes pH 7.4, 150 mM NaCl, 0.1% Triton, and 10% Glycerol), resuspended in Laemmli sample buffer, heated to 95°C for 10 minutes, resolved by electrophoresis on SDS-polyacrylamide gels, and immunoblotted to nitrocellulose using standard techniques (Harlow and Lane, 1988, Antibodies: A Laboratory Manual (Cold Spring Harbor, NY; CSH Laboratory Press)). Immunoblots were visualized using the ECL chemiluminescence system (Amersham).

20

Kinase Assays

To assay for MAP kinase activity, cells were lysed in freshly prepared lysis buffer (10 mM Tris HCl pH 7.5, 1% Triton X-100, 0.5% NP-40, 1 mM EDTA, 1 mM EGTA, 1 mM Na₃VO₄, 50 mM NaF, 1 mM PMSF, 40 mM PPI, 10 μg/ml aprotinin, and 10 μg/ml leupeptin) and MAP kinase was immunoprecipitated using 1 μg/ml anti-ERK2 antibodies from equal amounts of protein. After 4 washes with lysis buffer, and 2 with kinase buffer (20 mM Hepes pH 7.4, 10 mM MgCl₂), the immunoprecipitates were incubated with kinase buffer including 0.5 mg/ml myelin basic protein (MBP, Sigma), 10 mM ATP and 5 μCi [γ-³²P]ATP for 30 minutes at 30°C. The reactions were stopped by the addition of 2X sample loading buffer (Sambrook et al. 1989,

supra), resolved by 15% SDS-PAGE and visualized by autoradiography. Assays for activation of JNK MAP kinases were carried out exactly as described above, except that the immunoprecipitated JNK was incubated with 5 μ g purified
5 GST-c-Jun (Benn et al., 1996, supra) and 10 μ Ci [γ - 32 P]ATP, and resolved by 10% SDS-PAGE.

Src kinase assays were carried out essentially as described (Kypta et al., 1990, Cell 62:481-492). Briefly, NIH 3T3 or Chang cells were lysed in buffer containing 1% NP-40, 150 mM NaCl, 20 mM Tris HCl pH 8, 2.5 mM EDTA, 1 mM
10 Na_3VO_4 , 50 mM NaF, 1 mM PMSF, 10 μ g/ml aprotinin, and 10 μ g/ml leupeptin. Src and Fyn kinase activity were measured by immunoprecipitating either Src (1 μ g anti-Src antibodies) or Fyn (1 μ g/ml anti-Fyn antibodies) from equal amounts of cell lysate. The immunoprecipitates were washed four times with
15 lysis buffer, once with 20 mM Hepes pH 7.4 and once with kinase buffer (20 mM Hepes pH 7.4, 10 mM MnCl_2). Immunoprecipitates were then resuspended in kinase buffer containing 0.2 μ g acid denatured enolase (rabbit muscle; Sigma) and 20 μ Ci [γ - 32 P]ATP or 1 μ g Src-specific substrate
20 peptide (Upstate Biotechnology), 10 mM ATP and 10 μ Ci [γ - 32 P]ATP and incubated at 30°C for 30 minutes. Kinase reactions containing enolase were stopped with 2X sample loading buffer, heated for 10 minutes at 95°C, and resolved by 10% SDS-PAGE. For the reactions containing the Src
25 peptide substrate, TCA was added to a final concentration of 20%, an aliquot was spotted onto phosphocellulose paper (UBI), the filters were washed 3 X 5 minutes in 0.75% phosphoric acid and once in acetone, dried and analyzed by scintillation counting. Triton X-100 fractionation prior to Src kinase assays were carried out as described (Clark et
30 al., 1993, Moll. Cell Biol. 13:1863-1871).

To assay for p34 cdc2 kinase activity, Chang cells were lysed in 1% Triton, 20 mM Tris HCl pH 8, 10 mM EDTA, 5mM

MgCl₂, 1 mM Na₃VO₄, 50 mM NaF, 1 mM PMSF, 10 μg/ml aprotinin, and 10 μg/ml leupeptin. p34 cdc2 protein was immunoprecipitated from equal amounts of cell extract, washed four times with lysis buffer, twice with kinase buffer (50 mM Hepes pH 7, 10 mM MgCl₂, 5 mM MnCl₂, 4 mM dithiothreitol [DTT]), resuspended in kinase buffer containing 50 μg/ml Histone H1 (Boehringer-Mannheim) and 5 μCi [γ-³²P]ATP, and incubated at 30°C for 30 minutes. After stopping the reactions with sample loading buffer, the reactions were resolved by 12% SDS-PAGE, visualized by autoradiography, and quantitated by PhosphorImager analysis as described above.

Preparation of Nuclear Extracts

Nuclear extracts were prepared according to a modified Dignam protocol (Su and Schneider, 1996). Cell pellets were resuspended in 250 μl of cold Buffer A (10 mM Hepes pH 7.9, 1.5 mM MgCl₂, 10 mM KCl, 0.5 mM DTT, 1 mM PMSF, 10 μg/ml leupeptin and 10 μg/ml aprotinin), swollen on ice for 10 min, vortexed for 10 sec, and centrifuged for 10 sec at 12,000 x g. Nuclear pellets were resuspended in 30 μl cold buffer C (20 mM Hepes pH 7.9, 25% glycerol, 420 mM NaCl 1.5 mM MgCl₂, 0.2 mM EDTA, 1 mM DTT, 1 mM PMSF, 10 μg/ml leupeptin and 10 μg/ml aprotinin), incubated on ice for 20 min and nuclear extracts obtained by centrifugation at 12,000 x g for 2 min at 4°C.

25

6.2 RESULTS

To determine whether HBx induces an increase in the tyrosine kinase activity of the Src family of kinases, serum-starved NIH 3T3 cells were infected with recombinant adenoviruses (Ad) expressing either wild-type HBx (AdCMV-X) or a mutant HBx gene which transcribes a mRNA lacking all AUGs, and is therefore not translated (AdCMV-Xo). These vectors allow for rapid delivery of HBx genes and do not

30

activate Ras in the absence of HBx (Benn & Schneider, 1994, supra). c-Src was immunoprecipitated from an equal amount of extracted cell protein 8 hours post-infection (p.i.), the immune complex pellets were washed extensively, and an *in vitro* kinase assay was carried out using immunoprecipitated c-Src (auto-phosphorylation) and enolase (trans-phosphorylation) as substrates (Figure 1A). Expression of HBx, but not HBxo, induced a specific 3-4 fold increase in the ability of Src to undergo both auto- and trans-phosphorylation. Stimulation of cells with PDGF for 5 minutes, a prototypic Src induction protocol, induced a similar 3-4 fold activation. The observed PDGF-induced activation of c-Src is comparable to that reported by other groups (Gould and Hunter, 1988, *Mol. Cell Biol.* 8:3345-3356, Kypka et al., 1990, supra) and indicates that HBx expression induced activation of c-Src to an extent similar to that detected following treatment with PDGF. Identical trans-phosphorylation results were observed in Chang cells (Figure 1D). To determine whether HBx also activates other Src family members, serum-starved NIH 3T3 cells (Figure 1B) were infected with AdCMV-X or AdCMV-Xo viruses, c-Fyn was immunoprecipitated from equal amounts of cell lysates, and the immunoprecipitated pellets were subjected to an *in vitro* auto-phosphorylation and trans-phosphorylation kinase assay with the substrate enolase (Figure 1B). Expression of HBx induced an increase in the tyrosine kinase activity of c-Fyn, comparable to activation of Fyn following PDGF (Figure 1B) and insulin stimulation (Figure 1E), which was not observed in the HBxo-expressing cells. Identical trans-phosphorylation results were observed in Chang cells (Figure 1E). These data demonstrate that HBx induces the activation of different members of the Src family of nonreceptor tyrosine kinases.

To ensure that the activation of Src was not an unforeseen consequence of infection with the recombinant Ads, and to determine whether HBx activates c-Src in the cytoplasm or nucleus, Chang cells were transfected with plasmids expressing either wild-type HBx, mutant HBxo, or the nuclear-targeted variant HBxNLS. HBx-NLS contains a functional nuclear localization signal which causes the protein to become exclusively relocated to the nucleus. Following transfection, cells were serum-starved for 24 h, c-Src was immunoprecipitated from equal amounts of cell lysate, and an *in vitro* kinase assay was carried out using enolase as a trans-phosphorylation substrate (Figure 1C). Src kinase activity was strongly activated in serum-starved cells expressing wild-type HBx for 24-36 h, but not in those expressing HBxo or HBx-NLS. These results are consistent with the ability of HBx to activate Ras signaling cascades only in the cytoplasm. They also confirm that it is HBx which activates c-Src, and that a cytoplasmic location is essential for HBx activity. The plasmids all synthesize HBx mRNAs containing the adenoviral late 5' noncoding region (tripartite leader), which permits efficient translation under serum-free conditions as used here (reviewed in Schneider, 1996, in *Translational Regulation*, Cold Spring Harbor Press, Cold Spring Harbor). Thus, these data demonstrate that HBx is capable of activating c-Src without serum for a prolonged period of time (24-36 h), and only when located in the cytoplasm. It should be noted that conditioned media from cells expressing HBx for 8 h did not activate c-Src in control cells, excluding the possibility that HBx acts by inducing secretion of autocrine factors.

30

HBx activation of Ras signaling requires Src family kinases.

Src family kinases were shown to be negatively regulated by phosphorylation of a regulatory carboxy-terminal tyrosine by the action of the protein tyrosine kinase C-terminal Src kinase (Csk). Csk specifically phosphorylates the carboxy-terminal tyrosine and returns Src to an inactive state. Therefore, overexpression of Csk can be used as a mechanism to negatively regulate Src and potentially inhibit the ability of HBx to activate the Ras cascade. To test whether HBx activation of the Src family of kinases is required for stimulation of Ras signaling by HBx, Chang cells were transfected at low density with plasmids expressing HBxo, HBx or HBx and Csk, serum-starved for 20 hours, 2% serum restored for 2 hours to permit protein synthesis, and activation of immunoprecipitated MAP kinase at the end of the Ras-MAP kinase cascade assayed by incubation with myelin basic protein (MBP) and [γ - 32 P]ATP (Figure 2). Expression of HBx induces a strong activation in the ability of MAP kinase to phosphorylate MBP that is abolished upon co-transfection of HBx with Csk. In contrast, overexpression of Csk only slightly impaired activation of MAP kinase by insulin, indicating that expression of Csk did not generally impair activation of the Ras signaling cascade. Thus, these data clearly demonstrate that HBx requires activation of the Src family of kinases for induction of the Ras signaling cascade.

The results presented in Figure 2A were obtained in the presence of low serum (2% CS), therefore it was necessary to rule out the possibility that HBx only required Src activation for stimulation of ERK activity in the presence of serum. Furthermore, experiments were conducted to determine whether HBx, without the presence of serum, continued to stimulate ERK MAP kinase activity through the prolonged induction of Src family kinases observed in Figure 2A. Chang cells were transfected with plasmids expressing HBxo, HBx, or

HBx with Csk, kinase-inactive Src (pCMVSK⁻), or
kinase-inactive Fyn (pCMVFK⁻). Kinase-inactive forms of both
Src and Fyn have been shown to act in a dominant inhibitory
fashion to impair activation of downstream Src targets and
5 are considered dominant-negative proteins (Rusanescu et al.,
1995, supra; Twamley-Stein et al., 1993, supra). Cells were
serum-starved in 0.5% CS for 24 hours following transfection,
and activation of ERK2 assayed by immunoprecipitation and
incubation with MBP and [γ -³²P]ATP. ERK2 immunoprecipitates
10 obtained from serum-starved cells expressing HBx for 36 hours
displayed a 5 to 10 fold increase in kinase activity.
Moreover, co-expression of HBx with Csk, SrcK⁻, and FynK⁻
impaired activation of ERK2, although activation of ERK2 was
inhibited to a lesser extent by SrcK⁻. Although these
results indicate that both Src and Fyn play a role in
15 activation ERK by HBx, the precise contribution that each
makes to stimulation of Ras signaling by HBx was not
characterized. Nevertheless, these results demonstrate that
HBx stimulates a prolonged activation of ERK2 via stimulation
of Src family kinases, and does so without a requirement for
20 serum.

A more distant family member of the MAP kinase family,
Jun-NH₂ terminal kinase (JNK) (Derijard et al., 1994),
greatly enhances the transcriptional activity of c-Jun
through binding to the c-Jun transactivation domain and
phosphorylating it at Ser-63 and Ser-73 (Hibi et al., 1993,
25 Genes Devel. 7:2135-2148). Recent work from our lab has
demonstrated that HBx stimulates the activity of the JNK
members of the MAP kinase family, through the Ras pathway
(Benn et al., 1996 J. Virol 70:4978-4985). Experiments were
carried out to determine whether HBx induction of JNK
30 activity occurs through activation of the Src family kinases
similar to HBx stimulation of the ERKs. Chang cells were
transfected with plasmids expressing HBxo, HBx, HBx and

dominant-negative Src (SrcK⁻), or HBx and dominant-negative Fyn (FynK⁻), and serum-starved for 24 hours. JNK activity was assayed by immunoprecipitating JNK protein from equal amounts of cell lysate and incubating it with purified
5 GST-c-Jun (1-223), which contains the N-terminal site of phosphorylation, and [γ -³²P]ATP. Expression of HBx induced a strong (~20 fold), sustained activation of JNK which was not observed with HBxo expression, and was completely inhibited by co-expression with the dominant-negative Src and Fyn
10 plasmids. These results illustrate that sustained induction of JNK activity by HBx occurred through activation of Src family kinases. In contrast, expression of HBx did not alter the steady-state level of JNK1 protein. Taken together, the results confirm that an essential process in HBx induction of the MAP kinase family is its activation of Src family members
15 which in turn activates Ras and downstream ERK and JNK MAP kinases.

HBx transactivation of transcription factor AP-1 involves essential activation of Src kinases.

20 Here it is shown that HBx activation of AP-1 DNA binding activity and AP-1 directed transcription, both involve an essential requirement for HBx activation of Src tyrosine kinases. Chang cells were transfected with plasmids expressing HBx or HBxo proteins, with or without
25 cotransfection of Csk, serum starved and nuclear extracts prepared for electrophoretic mobility shift assay (EMSA) using equal amounts of nuclear extracts and a ³²P-labeled dsDNA probe containing one AP-1 binding site, as described (Benn et al., 1996 *supra*). Cotransfection of increasing
30 amounts of the Csk expressing plasmid suppressed HBx induction of AP-1 DNA binding complexes in a titratable manner (Figure 3A). Finally, cells were cotransfected with plasmids expressing HBx or HBxo and an AP-1 dependent

luciferase reporter under the control of a minimal promoter, with and without a Csk expression plasmid (Figure 3B). Co-expression with Csk fully blocked HBx induced activation of AP-1 dependent transcription, again demonstrating that
5 activation of Src kinases is essential for transcriptional transactivation by HBx.

HBx induces N-Myc and stimulates cyclin-dependent kinases by activating c-Src

10 The following experiments demonstrate that HBx induction of N-Myc requires HBx activation of Src family kinases. Chang cells were transfected with plasmids expressing HBxo, HBx, Csk, SrcK⁻, FynK⁻, and the dominant-negative Ras (RasN17). The Ras N17 mutant was initially shown to preferentially bind GDP *in vitro* and to inhibit cellular
15 proliferation when stably expressed in NIH 3T3 cells (Feig et al., 1988, Moll. Cell Biol. 8:3235-3243). Transfected cells were serum-starved for 20 hours, nuclear extracts prepared, equal amounts of protein resolved by SDS-PAGE and immunoblotted with anti-N-Myc antibodies. Expression of
20 transfected HBx elevates the amount of N-Myc protein 3-4 fold. Co-expression of HBx with Csk, and to a lesser extent the dominant-negative SrcK⁻, impaired the increase in N-Myc protein by HBx. The inability of dominant-negative Ras or Fyn proteins to block Src activation is consistent with the report by Barone and Courtneidge (1995, supra).
25 Co-expression of HBx with the dominant-negative Ras had no effect on induction of N-Myc by HBx, and neither did the expression of dominant-negative Fyn protein. Whether other Src family members (e.g. Yes or Lyn) are involved in N-Myc induction by HBx was not tested in these experiments. These
30 data indicate that HBx elevation of N-Myc occurs through a c-Src-dependent, Ras-independent pathway.

7. **EXAMPLE: THE HBx PROTEIN PROMOTES PROLONGED
WHV REPLICATION IN CHANG CELLS**

Conflicting results have been reported as to the requirement for HBx during the viral life cycle in cultured cells. While one group found no dependence on HBx for either the synthesis of HBV DNA or virion export in Huh7 and HepG2 cells (Blum et al., 1992 J. Virol. 66: 1223-1227), others have reported that genomic DNAs containing a wild-type HBx protein produce elevated levels of replicated viral DNA (Yaginuma et al., 1987 Proc. Natl. Acad. Sci. 84:2678-2682; Zoulim et al., 1994 J. Virol. 68:2026-2030). In order to determine the validity of studying the activities of HBx in the WHV system, the following analyses were conducted.

7.1 MATERIALS AND METHODS

15 Southern Blot Analysis of Viral Core-associated DNA

Chang cells were transfected with infectious WHV clones, (2 x 10 cm plates) were washed twice with PBS and lysed in 500 μ l lysis buffer/10 cm plate (20 mM Tris HCl pH 7.5, 1 mM EDTA, 100 mM NaCl, 0.5% NP-40) for 10-15 minutes. Lysates were centrifuged at 12,000 x g for 30 minutes, MgCl₂ was adjusted to 10 mM, DNaseI (Boehringer Mannheim) was added to 20 U/ml and the lysates were incubated at 37°C for 2 hours. Following DNase I treatment, the lysates were centrifuged for 5 min at 12,000 x g, layered onto a 20% sucrose cushion (300 μ l of 20% sucrose in 20 mM Tris HCl pH 8.0, 150 mM NaCl) and centrifuged for 178,000 x g for 3 hours at 20°C. The pellet was then resuspended in 250 μ l digestion buffer (10 mM Tris HCl pH 7.5, 10 mM EDTA, 1% SDS, 1 mg/ml proteinase K [Boehringer Mannheim]) and digested for 16-20 hours at 37°C. After extracting once with phenol and twice with phenol:CHCl₃, 100 μ l H₂O was added and the core-associated DNA precipitated using 0.2 M NaCl/2½ vol. ethanol and resuspended in 20 μ l TE (10 mM Tris HCl pH 7.5, 1 mM EDTA). 10 μ l

DNA/lane was electrophoresed through a 1.2% agarose gel and transferred to Duralon UV membrane (Stratagene). The filters were pre-hybridized at 42°C in 50% formamide, 5X SSC, 2.5X Denhardt's, 0.1% SDS, 1 mM EDTA, 5 mM NaH₂PO₄, and 100 µg/ml salmon sperm DNA for 3-24 hours, then hybridized in the same solution to a radiolabeled probe prepared from full length pWHV, HindIII digested and labeled to high specific activity with [α -³²P] dGTP and [α -³²P] dCTP. Hybridization was for 24-72 hours at 42°C (Sambrook et al., 1989, supra). The filters were washed at room temperature three times with 1X SSC, 0.1% SDS, three times at 50°C with 0.1X SSC, 0.1% SDS and autoradiographed for 1-2 days at -70°C.

Northern Analysis

Chang cells were washed twice with PBS and total RNA prepared according to the manufacturer's instructions (RNeasy mini kit; Qiagen). 15 µg total RNA per sample were resolved through a 1.2% formaldehyde-agarose gel (Sambrook et al., 1989, supra), transferred to Duralon UV membrane, and pre-hybridized in Quick-Hyb solution (Stratagene) at 65°C. Hybridization to ³²P-labeled full length WHV probe was carried out in the same solution at 65°C, washed as described above, and autoradiographed for 1 day.

7.2 RESULTS

To assess the requirement for the HBx protein during WHV replication in Chang cells, transfection experiments were carried out using either a replication-competent cDNA of WHV that expresses the wild-type WHx (wtWHV), or a replication-impaired cDNA which expresses a C-terminally deleted (inactive) WHx mutant (pXWHV). wtWHV and pCWHV encode a cDNA corresponding to the viral pregenomic RNA which is driven by the CMV promoter. While wtWHV directs the expression of infectious WHV in tissue culture cells and can initiate a

productive infection when implanted into the livers of susceptible woodchucks, pCWHV is non infectious in animals (Zoulim et al., 1994, supra). Chang cells were transfected at low confluency (see Methods) with plasmids encoding either wtWHV or pCWHV and the transfected cells were passaged when
5 confluent and propagated in culture for 11 days. To purify viral intracellular core-associated DNA, equal amounts of transfected cells were lysed in 0.5% NP-40, centrifuged through a 20% sucrose cushion, isolated viral cores were incubated with SDS and Proteinase K, and the viral core-associated DNA was purified by phenol extraction and ethanol
10 precipitation. The purified viral DNA was electrophoresed through a 1.2% agarose gel, transferred to nylon membrane, and a Southern blot analysis was carried out by hybridization of the viral DNA to a ³²P-labeled full-length genomic WHV DNA probe (Figure 4A). As compared to the almost undetectable
15 level of core associated-DNA purified from cells transfected with pCWHV (HBx mutant), the abundance of core-associated DNA isolated from wtWHV transfected cells was strongly enhanced by ~20-30 fold.

20 The hepadnavirus replication scheme discussed earlier dictates that the DNA purified from viral core particles can exist in several different forms which correspond to different stages of genomic maturation. These forms include abundant free minus strands, rare linear DNA duplexes and
25 substantial amounts of mature, relaxed duplex DNA species (Ganem, 1996 In Fields Virology, B.N. Fields, D.M. Knipe and P.M. Howley, eds. (Philadelphia:Lippincott Publishers) pp. 2703-2737). The presence of various forms of DNA within viral cores therefore reflects the presence of ongoing viral replication. The DNA isolated from wtWHV-expressing cells
30 appeared to contain both relaxed circular and single-stranded forms, indicating active replication. Furthermore, these results demonstrated that WHV was able to maintain a

replicative life cycle at least over the course of 11 days in Chang cells.

In comparison, HepG2 cells were also transfected with wt WHV, pCWHV, and pAUGWHV plasmids. pAUGWHV contains a
5 mutation of the first AUG to a UUG in the WHx ORF, rendering the X mRNA incapable of synthesizing WHx protein. Like pCWHV, pAUGWHV is non-infectious in animals (Zoulim et al., 1994). Transfected HepG2 cells were propagated for 14 days, the intracellular viral core-associated DNA was purified as
10 described above, and the viral DNA subjected to Southern blot hybridization to a ³²P-labeled full-length WHV genomic probe (Figure 4B).

The presence of the HBx protein enhanced the abundance of WHV core-associated DNA, although the upregulation of viral DNA synthesis by WHx (~5 fold) was not nearly as
15 pronounced as that observed in Chang cells (Figure 4A). To further confirm that the strong increase in the abundance of viral DNA replicative DNA intermediates in the presence of WHx is due solely from expression of the HBx protein, experiments were carried out to assess whether HBx protein
20 expressed *in trans* could complement the pCWHV defect in viral DNA synthesis shown in Figure 4A. Chang cells were co-transfected with either pCWHV and pCMV-HBx_o, or with pCWHV and pCMV-HBx, intracellular viral cores were isolated 3 days post-transfection, and the core-associated DNA analyzed by
25 Southern blot analysis as described above (Figure 5). Expression of HBx *in trans* complemented the pCWHV deficiency in viral DNA synthesis, although HBx *trans*-complementation did not entirely restore viral DNA synthesis to wild type levels (compare wt WHV in Figure 4A to Figure 5). This
30 discrepancy may result from a different level of expression of the HBx protein from pCMV-HBx as compared to expression from the wtWHV genomic DNA, or possibly unequal transfection efficiencies. Regardless, these results establish that HBx

can trans-complement pCWHV replication. These data also show that expression of the HBV HBx protein restored the ability of pCWHV to synthesize viral DNA intermediates, indicating that HBx and WHx are functionally interchangeable.

5

8. **EXAMPLE: HBx STIMULATES SRC-RAS SIGNALING
DURING IN VITRO VIRAL REPLICATION**

The following analysis was conducted to determine if HBx activates Src-Ras signaling when expressed in the context of in vitro viral replication (i.e., in tissue culture cells).

10

In these studies, HBV replication was supported in Chang cells with head-to-tail HBV dimers. Previous studies showed that transfection of either closed circular HBV DNA (Sureau et al., 1986 Cell 47:37-47) or HBV DNA in which two copies of the genome have been arranged in a head-to-tail dimer (HTD) can support *in vitro* production of viral proteins, synthesis of DNA replicative intermediates, and export of virions in permissive cells (Blum et al., 1992, J. Virol. 66:1223-1227; Yaginuma et al., 1987, Proc. Natl. Acad. Sci. 84:2678-2682). Due to the overlapping nature of the ORFs found in the circular genome of HBV, plasmids containing only one copy of the HBV genome interrupt viral coding regions, do not produce viral proteins and will not replicate (Dubois et al., 1980). These studies were conducted to directly determine whether HBx activates Ras signaling when synthesized from a genomic HBV HTD.

15

20

25

8.1 **MATERIALS AND METHODS**

The experiments were conducted using the materials and methods described in Sections 6.1 and 7.1 *infra*.

30

2025 RELEASE UNDER E.O. 14176

8.2 RESULTS

To determine whether WHx also requires activation of Src family kinases for stimulation of Ras signaling during ongoing viral replication, Chang cells were co-transfected with plasmids synthesizing wtWHV and dominant-negative Ras (RasDN), dominant-negative Src (SrcK⁻), or Csk proteins, and activation of MAP kinase was measured. As a control, Chang cells were also transfected with the WHx mutant plasmid, pCWHV. Eighteen hours post-transfection, cells were serum-starved for 24 hours, MAP kinase (ERK2) was immunoprecipitated from equal amount of cell lysate, and the immunoprecipitated protein was incubated with MBP and [γ -³²P]ATP (Figure 6). Consistent with our results described *infra*, wt WHV but not pCWHV strongly elevated the phosphorylation of MBP by MAP kinase. This indicates that the HBx protein stimulates Ras signaling during active viral replication. Furthermore, co-expression of wtWHV with either Ras or Src dominant-negative proteins, or with Csk protein, efficiently inhibited downstream activation of MAP kinase. These results indicate that the HBx protein activates a Src-Ras signaling cascade during active viral replication in cultured cells. Furthermore, co-expression of wtWHV with either the Ras or Src dominant-negative proteins, or with Csk protein, efficiently inhibited downstream activation of MAP kinase, indicating that the HBx protein activated a Src-Ras signaling cascade during *in vitro* viral replication.

9. EXAMPLE: WHV REQUIRES SRC FAMILY KINASES FOR IN VITRO REPLICATION

Given our observations described *infra* that HBx activates a Src-Ras signaling cascade during viral replication *in vitro*, and that the activation Src kinases by HBx mediated both Ras activation and cell cycle progression, experiments were next carried out to determine whether HBx

activation of Src is an essential function during viral replication.

9.1 MATERIALS AND METHODS

5 The experiments were conducted using the methods and materials described in Sections 6.1 and 7.1 *infra*.

9.2 RESULTS

10 Chang cells were co-transfected with plasmids encoding either wtWHV, pCWHV, wtWHV and RasDN, or wtWHV and Csk, intracellular core-associated viral DNA was isolated 3 days post-transfection, and purified viral DNA analyzed by Southern blot hybridization as described above (Figure 7). Consistent with the previous data, accumulation of viral DNA replicative intermediates was strongly enhanced in cells
15 expressing wtWHV, as compared to cells expressing pCWHV. Co-expression of RasDN protein with wtWHV had no detectable effect on viral replication, and viral DNA was synthesized at near wild-type levels. Expression of the RasDN protein impaired WHV activation of MAP kinase under these same
20 experimental conditions (Figure 7), indicating that the RasDN protein was able to function during WHV replication. However, these results illustrate the effects of one particular inhibitor of Ras and do not provide an explanation of the mechanism by which activation of Src kinase supports HBV replication. In sharp contrast, co-expression of Csk
25 with wtWHV completely abolished the ability of wtWHV to replicate to high levels. These results demonstrated that an essential component of the requirement of HBx during *in vitro* viral replication in Chang cells is its ability to activate Src signaling cascades, and that activation of Src family
30 kinases has a critical role during the viral replicative life cycle.

It was next addressed whether the stimulation of viral replication by WHx was a result of transcriptional transactivation of WHV genes by WHx. Chang cells were transfected with wtWHV or pCWHV plasmids. Total cellular RNA was isolated 4 days post-transfection, equal amounts of RNA were resolved by electrophoresis through a 1.2% formaldehyde-agarose gel, and the viral mRNAs were visualized by Northern blot analysis using a ³²P-labeled probe corresponding to the entire WHV genome (Figure 8, lanes 1 and 4). Steady-state levels of the genomic viral mRNAs was enhanced ~3-5 fold in Chang cells expressing wtWHV as compared to pCWHV. These data suggest that HBx protein induced only a moderate increase in the amount of viral mRNA, which cannot account for its striking stimulation of viral replication.

To assess whether HBx activation of a Src signaling cascade plays an essential role in transcriptional upregulation of the viral mRNAs, Chang cells were co-transfected with wtWHV and either RasDN or Csk plasmids, and the RNA visualized by Northern analysis (Figure 7, lanes 2 and 3). Co-expression of RasDN with wtWHV only slightly reduced the amount of the RNA species, while co-expression of wtWHV with Csk reduced the RNA level ~3-5 fold, to the level also observed by expression of pCWHV. To ensure that the decrease in synthesis of WHV RNA by Csk was not the unforeseen consequence of Csk inhibition of the CMV promoter (which drives synthesis of the WHV pregenomic RNA), control experiments assessing the effect of Csk on a CMV-βgal reporter were carried out. In comparison with expression of CMV-βgal alone, co-expression of Csk with CMV-βgal did not significantly alter expression of the βgal protein as measured by its β-galactosidase activity (Sambrook et al, 1989, supra). This control experiment indicates that expression of Csk does not generally inhibit transcription of the CMV promoter, and rules out a non-specific effect of Csk

on viral transcription. Therefore, these data suggest that the HBx protein moderately increases the abundance of all the viral transcripts through activation of the Src family of kinases. However, the WHx-induced increase in mRNA abundance
5 (~3-5 fold) is much less than the HBx-induced increase in viral DNA synthesis (~20-30 fold). Therefore, transcriptional transactivation by HBx does not appear to fully account for the augmentation of viral replication by the HBx protein. The stimulation of Src signaling cascades
10 by HBx must therefore promote WHV replication independent of the effect of WHx on viral transcription. These results illustrate that HBx activates a Src-Ras signaling cascade during viral replication *in vitro* which is essential for the host cell to sustain HBV replication.

15 10. **EXAMPLE: WHV REQUIRES SRC FAMILY OF
KINASES FOR IN VIVO REPLICATION**

The requirement for activation of Src kinase family members in replication of WHV can be determined in woodchuck infected livers in the following manner. A 2-5 year old
20 woodchuck is experimentally infected using a pooled serum from previous chronic carrier woodchucks. After 2 years of chronic infection, determined by WHsAg ELISA, the infected liver is surgically removed, the liver is perfused as described (Jacob et al., 1994, Exp. Cell Res. 212:42-48),
25 hepatocytes are dispersed by collagenase treatment and plated onto collagen coated dishes in L15 medium supplemented with 5% fetal calf serum, hydrocortisone and insulin. To introduce an inhibitor of Src kinases into primary hepatocytes, the Csk gene is cloned into the left-end of a
30 replication-defective adenovirus vector under the control of the CMV promoter, as described (Doria et al., 1995, EMBO J. 14:4747-4757). Adenovirus vectors infect primary cells and express trans-genes efficiently, whereas it is not possible

TOZT60 9005650

to transfect such cells at a high rate. Within several days of plating, cells are infected with the Csk-adenovirus vector, medium is replaced with L15 medium lacking insulin and containing reduced serum (between 0.5-2%). Cells are then harvested at 2 and 4 days after introduction of the vector. The medium can be assayed for levels of secreted WHV by ELISA for WHcAg and WHsAg. The level of virus replication can be assayed as described for Chang cells.

11. **EXAMPLE: PERSISTENT WHV REPLICATION IN
CELL CULTURE REQUIRES HBx PROTEIN**

In cultured cells HBV and WHV generally replicate very poorly in the absence of HBx during transient studies (Zoulim et al., 1994, J. Virol. 68:2026-2030; Yaginuma et al., 1987, Proc. Natl. Acad. Sci. 84:2678-2682; Nakatake et al., 1993, Virol. 195:305-314; Colgrove et al., 1989, J. Virol. 63:4019-4026; Lee et al., 1995, J. Biol. Chem. 270:31405-31412). To characterize the function of HBx in HBV replication we first examined whether HBx is required to promote or maintain viral replication in culture. Chang and HepG2 cells were transfected with wt or HBx(-) WHV genomes. Most studies were conducted with WHV to reduce the risk of infectious exposure. The level of active viral replication was examined in cells at 3 and 11 days post-transfection. To do so, viral core protein particles were isolated, which contain HBV DNA replication intermediates, ranging from immature ssDNAs to mature circular dsDNAs. Active viral replication was evident at both 3 days and 11 days following transfection of Chang and HepG2 cells with wtWHV. CWHV(HBx-) viral replication was very low at 3 days post-transfection (20-30 fold decreased) and almost undetectable by 11 days. Results in HepG2 cells were similar to those of less-differentiated Chang cells, although viral replication levels were ~5 fold lower on a per cell basis after consideration of transfection efficiencies.

HBx therefore stimulates and promotes persistent viral DNA replication in cultured cells. Cotransfection of an HBx expression plasmid with the CWHV(HBx-) mutant genome demonstrated that HBx expressed *in trans* complements its replication, demonstrating that poor replication resulted solely from the lack of HBx expression. The inability to fully *trans*-complement replication of the HBx mutant virus probably results from lower accumulation of HBx produced by the non-replicating plasmid.

10 12. CA²⁺ - PYK2 SIGNALING BY HBV HBX PROTEIN

The following experiments were conducted to address the question of whether HBx acts on the calcium-Pyk2 pathway, and if so, whether this is important for HBx stimulation of transcription and/or viral DNA replication. Three hepatic cell lines were transfected with vectors expressing HBx protein and transcription factor AP-1 dependent luciferase reporter (Figure 8A). HBx induced a 3 to 6 fold increase in AP-1 directed transcription in all cell lines. Cotransfection of a dominant-inhibiting Pyk2 known as PKM (*DiKic et al.*, 1996, Nature 383: 547-550), prevented activation of AP-1. Control studies showed that PKM decreased HBx expression from the CMV promoter by half. Since transfection of cells with 10 fold less HBx expression plasmid only reduced activation of AP-1 by 1.8 fold (Figure 8A), these data indicate that PKM acts by inhibiting HBx activity rather than expression.

To determine whether HBx activates Pyk2 as indicated by Y-402 phosphorylation (*DiKic et al.*, 1996, Nature 383: 547-550), HepG2 and Chang cells were transfected with vectors expressing HBx or vector alone, with or without PKM expression. HBx induced a 4 fold increased phosphorylation of Pyk2 at Y-402, comparable to that of TPA stimulation, without altering Pyk2 abundance (Figure 8B). HBx activation

of Pyk2 was likely constitutive, as it was sustained 48 h following transfection, the last time point tested. HBx induced phosphorylation of Pyk2 correlated with stimulation of Pyk2 protein kinase activity, as shown by immunoprecipitation of Pyk2 and in vitro Pyk2 autophosphorylation using $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ (Figure 8C). Compared to vector transfected cells, HBx induced 6 fold increased activation of Pyk2, as well as autophosphorylation of c-Src and Fyn Src kinases, which electrophoretically comigrate. To determine whether HBx activation of Pyk2 is essential for downstream stimulation of Src kinases, Chang cells were transfected with vectors expressing HBx and PKM or empty vector, Fyn was immunoprecipitated and tested for autophosphorylation activation by in vitro kinase assay. HBx induced a 5 fold increase in Fyn phosphorylation activity compared to vector alone, which was prevented by coexpression with PKM (Figure 8D), demonstrating that HBx activation of Pyk2 activates Src kinases.

Studies determined whether HBx acts on intracellular calcium to activate Pyk2. HBx transfected Chang cells showed 5 fold increased phosphorylation of Pyk2 at Y-402, similar to TPA stimulation (Figure 9A). Treatment with the cell permeable cytosolic calcium chelator BABTA-AM at $50\ \mu\text{M}$ (2 times IC_{50}) for 2 h prevented Pyk2 phosphorylation without altering Pyk2 levels (Figure 9A). HBx activation of Pyk2 therefore involves cytosolic calcium action. Studies next determined whether HBx acts on calcium channels in the endoplasmic reticulum (ER), mitochondria or plasma membrane (PM) for its activity. A low ($0.5\ \text{mM}$) concentration of EGTA was added to the culture medium for 2 h to block entrance of extracellular calcium (Zwick et al. 1999, J.B.C. 274: 20989-20996), or cells were treated with BAPTA-AM to block ER and mitochondrial calcium, or cyclosporin A (CsA) to block mitochondrial calcium function. EGTA had no effect whereas

099500550

Sub.B6

BAPTA-AM or GsA both prevented HBx activation of Pyk2, indicating that HBx acts on ER/mitochondrial calcium control. A high concentration of EGTA (3 mM) did not block TPA activation of Pyk2 phosphorylation (Zwick et al. 1999, J.B.C. 274: 20989-20996) (Figure 9C), but partially inhibited activation by HBx. Therefore, HBx acts on the control of ER/mitochondrial calcium, with low level entry of extracellular calcium, suggestive of constitutive cytosolic calcium alteration (Clapham 1997, Cell 80:259-268).

The requirement for cytosolic calcium and HBx activation of Pyk2 in HBV replication was examined. HepG2 cells were transfected with a 130% head-to-tail DNA copy of the HBV genome which replicates authentically in the livers of transgenic mice (Guidotti et al. 1995, J. Virol. 69: 6158-6169), in *Tupaia* hepatocytes in culture (Melegari et al., 1998, J. Virol. 72: 1737-1743), and in an HBx-dependent manner in HepG2 cells (Melegari et al., 1998, J. Virol. 72: 1737-1743). Expression of HBx was abolished by a targeted frameshift mutation (Melegari et al., 1998, J. Virol. 72: 1737-1743). HepG2 cells were transfected with vector alone, wild type HBV genomic DNA, or HBx(-) genomic DNA, cytoplasmic viral core particles, the structures in which viral DNA replication takes place, were isolated and the level of viral DNA replication was examined (Figure 9D). HBV DNA replication was reduced 20 fold in the absence of HBx expression, but recovered by cotransfection of HBx. Northern niRNA analysis demonstrated no reduction in HBV pregenomic (pg)RNA and HBsAg mRNAs in the absence of HBx (Figure 9D). Cotransfection of wild type HBV genomic DNA with PKM reduced viral DNA replication by 15 fold, similar to HBx(-) HBV samples, without altering viral niRNA levels (Figure 9E). These results demonstrate that HBx specifically promotes HBV DNA replication in a Pyk2-dependent manner.

Sub E

095506-0471

The requirement for cytosolic calcium in HBx-dependent viral replication was investigated. Cells transfected with wild type or HBx(-) HBV genomic DNA were treated for 4 d with 1 or 3 p.g/ml of CsA to block mitochondrial calcium channels. There was no evidence for CsA toxicity during treatment. CsA reduced HBV DNA replication in cytoplasmic core particles by 15 fold compared to untreated controls (Figure 10A), similar in magnitude to inhibition of Pyk2 or the absence of HBx expression. Northern mRNA analysis demonstrated a 2 fold reduction in pgRLNA and HBsAg mRNAs (Figure 10A). To determine whether inhibiting cytosolic calcium and Pyk2 activity inhibits HBV DNA replication, HepG2 cells were transfected with HBV genomic DNA and treated with CsA, or cotransfected with PKM. Cytosolic core particles were purified and incubated with [α -³²P]-dNTPs to examine endogenous HBV polymerase activity (Figure 10B). In untreated controls, predominantly full-length double-strand DNA products were produced, indicative of pgRNA reverse transcription and DNA-dependent DNA synthesis. PKM inhibition of Pyk2 or treatment of cells with CsA prevented DNA replication by 7 and 12 fold respectively. Treatment of HBV genome transfected cells with low levels of BAPTA-AM for 4 d impaired viral DNA replication by 10 fold without strongly reducing HBV mRNA levels (Figure 10C). Collectively, these data show that HBx activation of HBV reverse transcription and DNA replication involves alteration of cytosolic calcium and coupled activation of Pyk2. The requirement for cytosolic calcium in HBx transcriptional stimulation was investigated in HepG2 cells transfected with luciferase reporters controlled by transcription factor AP- 1 or CREB, with or without treatment of cells by CsA (Figure 10D). HBx activation of AP- 1 dependent transcription was impaired 2.5 fold by treatment of cells with 10 μ g/ml CsA. HBx stimulation of CREB-dependent transcription was resistant

SAB
B7

0955006 09101

with CsA blocked HBx activation of HBV DNA replication. Third, ionophoric agents that increase the level of cytoplasmic calcium functionally replace HBx in viral DNA replication. Thus, HBx acts on stored cytosolic calcium as a
5 fundamental activity for HBV replication.

The present invention is not to be limited in scope by the specific embodiments described which are intended as single illustrations of individual aspects of the invention, and functionally equivalent methods and components are within
10 the scope of the invention. Indeed various modifications of the invention, in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

15 Various references are cited herein, the disclosures of which are incorporated by reference in their entireties.

20

25

30

FOIA b 7 - D